

# A Two-Metal-Ion-Mediated Conformational Switching Pathway for HDV Ribozyme Activation

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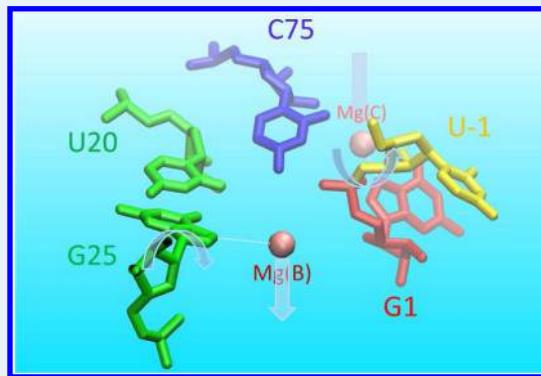
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## Supporting Information

**ABSTRACT:** RNA enzymes serve as a potentially powerful platform from which to design catalysts and engineer new biotechnology. A fundamental understanding of these systems provides insight to guide design. The hepatitis delta virus ribozyme (HDVr) is a small, self-cleaving RNA motif widely distributed in nature, which has served as a paradigm for understanding the basic principles of RNA catalysis. Nevertheless, questions remain regarding the precise roles of divalent metal ions and key nucleotides in catalysis. In an effort to establish a reaction mechanism model consistent with available experimental data, we utilize molecular dynamics simulations to explore different conformations and metal ion binding modes along the HDVr reaction path. Building upon recent crystallographic data, our results provide a dynamic model of the HDVr reaction mechanism involving a conformational switch between multiple noncanonical G25:U20 base pair conformations in the active site. These local nucleobase dynamics play an important role in catalysis by modulating the metal binding environments of two  $Mg^{2+}$  ions that support catalysis at different steps of the reaction pathway. The first ion plays a structural role by inducing a base pair flip necessary to obtain the catalytic fold in which C75 moves towards the scissile phosphate in the active site. Ejection of this ion then permits a second ion to bind elsewhere in the active site and facilitate nucleophile activation. The simulations collectively describe a mechanistic scenario that is consistent with currently available experimental data from crystallography, phosphorothioate substitutions, and chemical probing studies. Avenues for further experimental verification are suggested.



**KEYWORDS:** HDV, HDVr, QM/MM, ribozyme, reaction mechanism, conformation switching, metal ions

## INTRODUCTION

Biological enzymes are among the most complex, selective, and efficient known catalysts and have seen increasing use in industrial applications in comparison to chemical catalysts.<sup>1,2</sup> Proteins, with their diverse array of chemical functional groups, are extraordinarily efficient<sup>3</sup> and for a long time were thought to be the only biological catalysts. In the 1980s, however, it was discovered that RNA molecules could have catalytic activity, causing a veritable revolution in catalytic RNA research.<sup>4–8</sup> RNA molecules have several assets as a platform for creating catalysts, and the engineering of new biotechnology based on catalytic RNAs is widespread and ongoing.<sup>9</sup> Nonetheless, it remains an open chemical question as to how RNA, with limited available nucleotide building blocks that are largely chemically inert, can fold so as to adopt three-dimensional structures that are able to confer activity which rivals that of many protein enzymes.<sup>10,11</sup> Consequently, it is an active area of catalytic research to understand the possible mechanistic strategies RNA can employ. Toward this end, the structural and kinetic characterization of the

class of small nucleolytic RNA enzymes, or *ribozymes*, has been instrumental.<sup>12,13</sup>

The hepatitis delta virus ribozyme (HDVr), a small self-cleaving catalytic RNA motif, is a prototype nucleolytic ribozyme that catalyzes RNA 2'-O-transphosphorylation which involves an inline nucleophilic attack by U-1:O<sub>2'</sub> on the adjacent scissile phosphate and cleavage of the P–O<sub>5'</sub> bond of G1 to produce a 2',3'-cyclic phosphate and a 5' hydroxy terminus. Originally found as a satellite of the hepatitis B virus, where it plays a role in the viral life cycle,<sup>14–17</sup> HDV-like ribozyme sequences have subsequently been found in other prokaryotic and eukaryotic genomes (including that of humans).<sup>18,19</sup> The motif may be an important tool in as yet unrealized biological roles.<sup>20–23</sup> Extensive structural and biochemical evidence supports a mechanism involving general acid and metal ion catalytic modes.<sup>24,25</sup> However, there is still

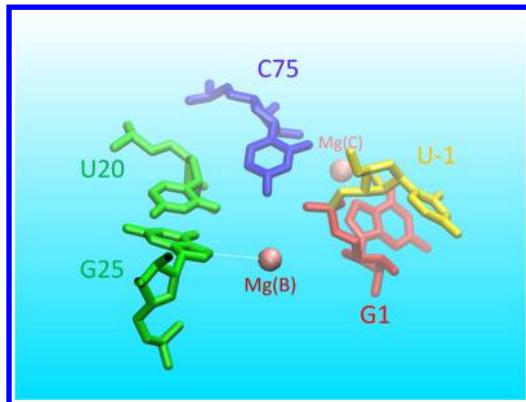
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no strong consensus on the exact role(s) of divalent metal ions in transition state stabilization or on the specific conformations required at different stages of the reaction (Figure 1).<sup>25,26</sup>



**Figure 1.** Active site of HDVr with two potential  $Mg^{2+}$  binding sites.

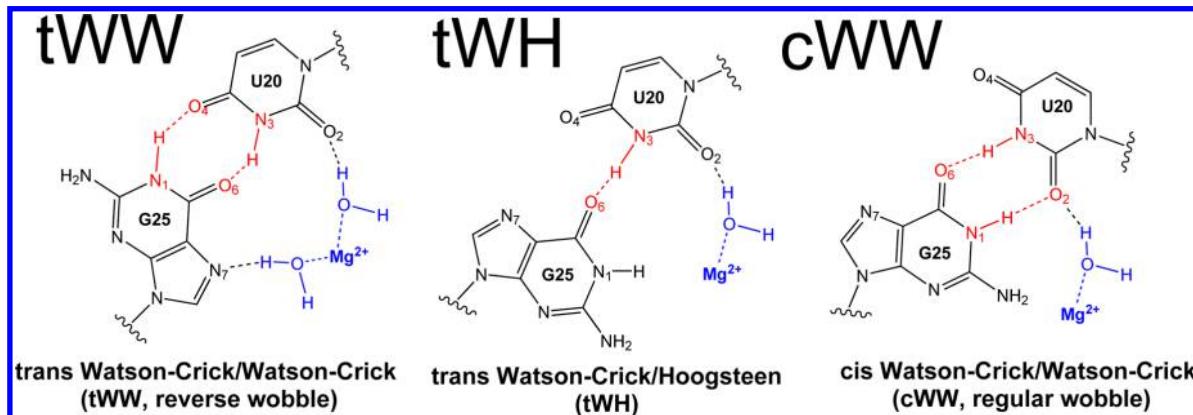
The first crystal structure of HDVr was reported in the cleaved (i.e., product) form<sup>27</sup> with an overall fold consistent with mutagenesis and chemical probing studies of the solution conformation.<sup>28,29</sup> In that structure, C75:N<sub>3</sub> is in a position to form a hydrogen bond with G1:O<sub>5'</sub> (the leaving group), suggesting that C75 acts as a general acid to stabilize the leaving group.<sup>30,31</sup> This hypothesis is supported by inactivation of the ribozyme by C75 mutation as well as other experimental approaches.<sup>30,32–36</sup> In particular, preactivation of the leaving group by phosphorothioate substitution renders the ribozyme insensitive to C75 mutation.<sup>31</sup>

Consistent with its role in acid catalysis, the protonation state of C75 is also critically important. It has been well established that the  $pK_a$  of C75 is shifted toward neutrality in order to act optimally as a general acid.<sup>30,33,34,36–38</sup> Nevertheless, the interactions that influence the protonation state of C75 along the reaction coordinate have not been resolved. Raman crystallography experiments reported that the  $pK_a$  of C75 is shifted to around 6.1–6.4 in the presence of different deactivated nucleophiles,<sup>35</sup> while NMR experiments have shown that the  $pK_a$  of C75 is not shifted significantly in the product and the reactant states in comparison to that in solution ( $pK_a \approx 4.5$ ).<sup>37</sup> As such, it is desirable to explore the

interactions between C75 and its neighboring residues at different stages of the reaction, especially as a function of protonation/deprotonation. Indeed, it has been suggested that C75 might be properly positioned for reaction only after the precursor is formed,<sup>39</sup> which was interpreted as supporting the role of protonated C75 as the general acid, but only near the transition state. Recently the protonation state of C75 has also been suggested to play a critical role in forming the inline conformation.<sup>40</sup>

Until recently, the crystallographic data from HDV ribozymes that have been inactivated by a C75U mutant was interpreted such that G25 and U20 form a weak trans Watson–Crick/Hoogsteen (tWH) base pair (Figure 2);<sup>41</sup> a divalent ion was also resolved bound to U75:O<sub>4'</sub>.<sup>42,43</sup> However, in structures of the cleaved product,<sup>28,29</sup> as well as a recently reported precleavage structure,<sup>44</sup> G25 and U20 are resolved as a “reverse wobble” pair (trans Watson–Crick/Watson–Crick, tWW) where G25:N<sub>7</sub> and G25:O<sub>6'</sub> may provide a preferred environment for  $Mg^{2+}$  binding (Figure 2).<sup>45,46</sup> Conversely, in a structure where divalent ions had been removed, the position of the G25 ribose is not resolved and the nearby C24 is not stacked on G25.<sup>42</sup> Recently, a C75U precleavage structure as well as a postcleavage structure were refined with new computational tools.<sup>47</sup> In these newly fitted structures G25 and U20 are also resolved in the “reverse wobble” (tWW) conformation. Finally, the L3 loop of HDVr (where the G25:U20 base pair is located) is believed to be highly flexible<sup>48</sup> and prone to misfolding.<sup>49</sup> Its conformation may also be metal ion dependent,<sup>50,51</sup> and chemical modification results suggest that G25 may be exposed to solvent during the course of the reaction.<sup>52</sup> Although the pair conformation may be flexible, both G25 and U20 are completely conserved across different species, implying they are critical for the reaction.<sup>19</sup>

Regarding the active conformation, on the basis of experimentally observed stereochemical inversion of the phosphorus center<sup>53–55</sup> lack of isomerization (RNA migration) products, it is generally accepted that small ribozymes require inline conformations to facilitate the  $S_N2$ -like mechanism of cleavage,<sup>53–56</sup> although the possibility of an alternative mechanism involving pseudorotation cannot be definitively ruled out.<sup>57</sup> Nevertheless, the inline conformation necessary for the HDVr reaction has never been observed in any reported crystal structure to date. Indeed, in the precleavage structures of the HDVr the inline attacking angles are less than 90°.<sup>42,44</sup> Although it has been



**Figure 2.** Three conformations of the G25:U20 pair in the HDVr reaction. Key hydrogen bonds distinguishing the three conformations are colored red: (1) tWW, G25:N<sub>1</sub>/U20:O<sub>4'</sub> and U20:N<sub>3</sub>/G25:O<sub>6'</sub>; (2) tWH, U20:N<sub>3</sub>/G25:O<sub>6'</sub> only; (3) cWW, U20:N<sub>3</sub>/G25:O<sub>6'</sub> and U20:O<sub>2'</sub>/G25:N<sub>1</sub>. Possible  $Mg^{2+}$  binding modes near G25:U20 are also shown in blue.

suggested that there may be some space for U-1 to undergo a rigid rotation to reach the necessary orientation,<sup>42</sup> such a rotation has not yet been directly tested experimentally.<sup>24</sup> A recently reported precleavage structure shows U-1 to be unresolved and hence does not provide direct structural evidence for an inline conformation, although it has been suggested that the HDVr inline conformation could be modeled on the basis of an active Hammerhead ribozyme structure.<sup>44,58</sup> Our previous simulations revealed a spontaneous rotation of U-1 that led to an inline conformation, apparently driven by deprotonation of U-1:O<sub>2'</sub>.<sup>59</sup>

Although significant constraints have been established, no clear “best” representative picture of the active site or the reaction path has yet emerged. Indeed, experimental observations seem to strongly suggest that the reaction is highly coupled with conformational switching events and metal ion binding modes.<sup>39,40,42,60</sup> As such, a key position in this work (and one not without precedent<sup>39,60</sup>) is that a single, static conformational description of the HDVr active site is insufficient to capture the complete mechanism.

A related unanswered question is the role(s) of the metal ion(s) in the HDVr reaction. Although not absolutely required for catalysis, the presence of divalent metal ions at millimolar levels significantly enhances the HDVr reactivity.<sup>16,28,61–63</sup> There are vast experimental data regarding the active site Mg<sup>2+</sup>, and it is believed that there is at least one hydrated Mg<sup>2+</sup> ion near the active site.<sup>26,30,42,64</sup> Some data have been interpreted as indicating two Mg<sup>2+</sup> ions in the active site,<sup>23,43,65,66</sup> and it has been suggested that the active site Mg<sup>2+</sup> ion(s) may play two types of roles.<sup>25,32,66,67</sup> The first is structural; the Mg<sup>2+</sup> ion binding facilitates the formation of the active conformation.<sup>32</sup> The second is chemical; an Mg<sup>2+</sup> bound hydroxide acts as a base extracting to activate the 2'-OH group of U-1,<sup>30,32,68</sup> or Mg<sup>2+</sup> could act in a Lewis acid role to lower the pK<sub>a</sub> of the O<sub>2'</sub> nucleophile.<sup>44,66</sup> Nevertheless, most recent mechanistic models have focused heavily on “the catalytically critical divalent metal ion”.<sup>36,42,46,65,66,68–72</sup>

Furthermore, it has been shown that Co(NH<sub>3</sub>)<sub>6</sub><sup>3+</sup> (which can compete with Mg<sup>2+</sup> in binding but cannot form inner-sphere coordination interactions) inhibits HDVr activity.<sup>30,70</sup> The active site Mg<sup>2+</sup> ion has been shown to directly interact with critical active site residues,<sup>65,73</sup> and modification of the linkage at the scissile phosphate can alter metal ion preference.<sup>62,74</sup> A recent crystal structure of the HDVr in the precleavage state (using a deoxy substrate), combined with molecular modeling, led to the hypothesis that the Mg<sup>2+</sup> directly coordinates U-1:O<sub>2'</sub> as well as the G1:*pro-R<sub>P</sub>* oxygen.<sup>44</sup> Thio substitution of the *pro-R<sub>P</sub>* oxygen of the scissile phosphate has been utilized to explore possible direct Mg<sup>2+</sup> binding, but no direct rescue effect from Mn<sup>2+</sup> was observed.<sup>75,76</sup> However, in recent studies an R<sub>P</sub> thio substitution was observed to result in two slow reaction phases, and an “unconventional” rescue of the R<sub>P</sub> thio effect was observed in which the rescuing ion, Cd<sup>2+</sup>, inhibits the wild-type but not the *pro-R<sub>P</sub>* thio-substituted ribozyme.<sup>71,72</sup> Large *pro-R<sub>P</sub>* thio effects are also observed in reactions with monovalent ions alone; however, the thio effects at the *pro-S<sub>P</sub>* position result in faster reaction rates (termed inverse thio effects). The pH profiles are very different: bell-shaped profiles are found with native Mg<sup>2+</sup> and other small divalent ions<sup>23,72</sup> but are inverted with large divalent ions or monovalent ions.<sup>30,32,36,72,77</sup> Additionally, a proton inventory of 1 is observed in monovalent ions while an inventory of 2 is observed

for reactions containing Mg<sup>2+</sup> consistent with participation of a metal hydrate in catalysis.<sup>78</sup>

Although detailed models of the roles of metals in the HDV mechanism have been proposed on the basis of extensive biochemical data, important ambiguities remain. A consistent description of the HDVr catalytic mechanism thus appears to be precluded by (1) various conformations of the active site (Figure 1) in the available structures and (2) different proposals regarding the role of the active site Mg<sup>2+</sup>. Similar ambiguities are inherent in many ribozymes and protein metallophosphoryltransferases. A recent simulation work suggests that crystal structures may represent trapped intermediates along the reaction coordinate.<sup>40</sup> A long-term molecular dynamics simulation can serve as a powerful tool complementing experimental observations to establish a hypothesis that consolidates all available empirical evidence.<sup>40,59,66,79–83</sup> Recent QM/MM simulations<sup>68,71,72,82,84</sup> starting from a proposed starting structural model of the active site<sup>44</sup> have delivered a detailed, even quantitative, picture of several reaction pathways in the HDVr. However, at the same time, these QM/MM simulations fail to explain most qualitative observations: e.g., the comparison of reaction barriers in the presence and absence of Mg<sup>2+</sup>.<sup>84</sup> In order to explore additional plausible scenarios of the HDVr reaction, here we significantly extend the scope of past simulation studies to include various starting conformations, C75 protonation states, and Mg<sup>2+</sup> binding modes at different stages of the HDVr reaction. The results inspire a novel, dynamic view of the HDVr reaction where conformational events (especially changes in G25:U20 base pairing), metal ion binding modes, the C75 protonation state, and chemical steps are all highly coupled.

## SIMULATION SETUP

This work greatly extends the scope of our previous simulations<sup>59</sup> with a consistent protocol. The extension of this work is briefly described below. Detailed description of the simulation setup and protocols can be found in the Supporting Information.

The Amber<sup>85,86</sup> parm99 force field with the  $\alpha/\gamma$  corrections for nucleic acids, which has been demonstrated to significantly improve nucleic acid side chain conformations,<sup>87</sup> was employed. The parameters of Mg<sup>2+</sup> specifically optimized for RNA binding were utilized.<sup>88</sup> In the present study, the purpose is to identify plausible catalytically active states (i.e., conformations, metal ion binding modes, and protonation states) along the reaction coordinate that can be used as a departure point for further investigation of the chemical steps of the reaction with more sophisticated QM/MM models.

More than 30 long-term ( $\geq 300$  ns each and roughly 10  $\mu$ s in total) molecular dynamics simulations at different stages along the HDVr reaction coordinate were performed with different starting conditions: the G25:U20 base pair in either its tWW or tWH forms<sup>41</sup> (Figure 2), C75 protonated or neutral, and Mg<sup>2+</sup> ion(s) in different binding modes.

**Mg<sup>2+</sup> Binding Sites.** It has been suggested that there are two possible Mg<sup>2+</sup> binding sites, one near G1:N<sub>7</sub><sup>43,58,65,70,89</sup> and another near the active site phosphate (possibly bound to U75:O<sub>6</sub> in the case of C75U mutants)<sup>43</sup> or near G25:N<sub>7</sub> in other structures.<sup>43,46,81</sup> In accordance with the previously reported notation,<sup>43</sup> the binding site near G1:N<sub>7</sub> is defined as the “C-site” and the site near U75:O<sub>6</sub>/G25:N<sub>7</sub> as the “B-site”. In all simulations reported here, the B-site initial position was either taken directly from the crystal structure (PDB: 1VC7)

when G25:U20 was in the tWH form or modeled by placing a Mg<sup>2+</sup> ion 4.0 Å from G25:N<sub>7</sub> when G25:U20 was in its reverse wobble form. The C-site initial position was at 2.0 Å from G1:N<sub>7</sub>, as suggested in other work.<sup>43,65</sup>

**G25:U20 Base Pairing Modes.** There are two types of G25:U20 base pair conformations in the reported crystal structures (**Figure 2**): trans Watson–Crick/Hoogsteen (tWH)<sup>42,43</sup> and the unusual trans Watson–Crick/Watson–Crick wobble (tWW, also referred to as a reverse wobble).<sup>28,29,44,47</sup> In order to explore these two possible G25:U20 base pairings, initial structures with different G25:U20 conformations were used. The initial starting positions of all other residues were the same as in our previous work,<sup>59</sup> which was derived from a C75U crystal structure (1VC7). To our knowledge, there is no fully resolved U-1 residue in any reported structure with G25:U20 in the tWW form. Hence, the following procedure was taken to build the initial structures with G25:U20 in the tWW form. After 10 ns of solvent and ion equilibration, the structures with G25:U20 in the tWH form were modified by rotating the G25 glycosidic bond torsion (O<sub>4'</sub>–C<sub>1'</sub>–N<sub>9</sub>–C<sub>4</sub>) to 67.5°. In this syn conformation two hydrogen bonds were nearly formed in the expected tWW pattern (G25:N<sub>1</sub>–U20:O<sub>4</sub> and G25:O<sub>6</sub>–U20:N<sub>3</sub>). This was followed by an additional 1 ns equilibration of solvent and ions. The resulting structures underwent further relaxation with two hydrogen bonds (G25:N<sub>1</sub>–U20:O<sub>4</sub> and G25:O<sub>6</sub>–U20:N<sub>3</sub>) restrained with 20 kcal/mol·Å<sup>2</sup> harmonic potentials for 10 ns to ensure the proper response and relaxation of the surrounding residues. The final structures then served as the starting structures for the reported MD simulations.

**C75 Protonation States and Stages along the Reaction Coordinate.** C75 in both its neutral and protonated forms were used in the simulations as reported previously.<sup>59</sup> In addition, simulations were performed at different stages of the reaction: reactant state, activated precursor state, early transition state mimic, late transition state mimic, and product state.

**Simulations Performed.** We use the following notations for Mg<sup>2+</sup> binding sites: M<sub>B</sub> stands for a Mg<sup>2+</sup> placed at the B-site initially and M<sub>C</sub> at the C-site. For G25:U20 base pairing modes, “tWH” stands for initial structures in the tWH pair form and “tWW” in the tWW (reverse wobble) form. For C75 protonation states, C75° stands for neutral C75, while C75<sup>+</sup> stands for protonated C75. Different stages of the reaction are marked as reactant state (RT), activated precursor state (dRT), early transition state mimic (ETS), late transition state mimic (LTS), and product state (PD). All simulations are given in the tables in the **Results and Discussion**.

## ■ RESULTS AND DISCUSSION

**B-Site Mg<sup>2+</sup> Ion Stabilization of the tWW Conformation That Brings C75 Close to the Scissile Phosphate, Resulting in a Downshift of the C75 pK<sub>a</sub>.** **Table 1** gives key geometric parameters characterizing the base pairing of G25:U20, the Mg<sup>2+</sup> binding of the B- and C-sites, and the position of C75 in reactant state simulations under various conditions. The glycosidic bond torsion angle,  $\chi$ , of G25 is an indicator of the *syn/anti* conformation of G25. Distances from C75:N<sub>3</sub> to both G1:O<sub>1P</sub> and G1:O<sub>2P</sub> were calculated and the average of their reciprocal values are shown as a measure of their electrostatic interaction ( $\langle(1/r_1) + (1/r_2)\rangle$  in all tables).

**Table 1. Key Geometric Parameters for B-Site Formation in Reactant State Simulations (RT\_C75°)<sup>a</sup>**

simulation	hydrogen bond			distance to C75:N <sub>3</sub>						distance to Mg <sup>2+</sup> (C)			
	G25:N <sub>1</sub> /U20:O <sub>4</sub>	G25:N <sub>6</sub> /G25:O <sub>6</sub>	U20:O <sub>2</sub> /G25:N <sub>1</sub>	G25: $\chi$	G1:O <sub>1P</sub>	G1:O <sub>2P</sub>	$\langle(1/r_1) + (1/r_2)\rangle$	U-1:O <sub>2</sub>	G1:O <sub>2P</sub>	G25:N <sub>7</sub>	U20:O <sub>2</sub>	U-1:O <sub>2</sub>	G1:N <sub>7</sub>
tWW_RT_C75°M <sub>B</sub>	85.8	76.0	75.5(14.0)	5.86(1.34)	5.09(1.62)	0.40	7.95(1.10)	4.49(1.36)	4.90(0.75)	4.57(0.52)			
tWW_RT_C75°M <sub>C</sub>	58.0	61.7	4.3	151.4(47.0)	4.55(0.66)	6.62(1.16)	0.38	12.45(4.10)	11.32(3.88)	16.51(3.29)	14.93(2.44)	12.45(4.10)	7.41(4.50)
tWW_RT_C75°	19.9	20.7	16.0	-169.2(11.3)	5.54(0.68)	4.84(0.63)	0.39						
tWW_RT_C75°_fix	83.8	87.7		-167.1(16.6)	4.59(0.65)	4.36(0.78)	0.46						
tWW_RT_C75°M <sub>B</sub>				-77.4(13.9)	10.44(0.98)	9.74(0.92)	0.20	4.43(0.73)	1.87(0.04)	7.59(0.34)	4.83(1.12)		
tWW_RT_C75°M <sub>C</sub>				-69.0(8.7)	4.10(1.02)	4.43(0.78)	0.49						
tWW_RT_C75°				-50.0(10.7)	5.70(0.84)	7.58(1.09)	0.31				7.82(2.77)	5.62(3.90)	

<sup>a</sup>Notations used: initial G25:U20 base pairing: tWW (reverse wobble) and tWH (**Figure 2**); M<sub>B</sub>, Mg<sup>2+</sup> initially placed at the B-site (near G25:N<sub>7</sub>); M<sub>C</sub>, Mg<sup>2+</sup> initially placed at the C-site (near G1:N<sub>7</sub>); C75°, C75 is neutral; RT, reactant state. Entries are calculated from a sampling of the range 50–300 ns from the 300 ns trajectory for each simulation condition with a sampling free frequency of 100 ps, resulting in 2500 data points for every simulation. Distances are given in Å, and hydrogen bond occupations (given as percentages) are defined as the percentage of the snapshots in which the distance between the proton donor and the proton acceptor  $r \leq 3.5$  Å and the H-bond angle  $\theta \geq 150^\circ$ . The numbers are average values, and corresponding standard deviations are given in parentheses. The results suggest the following: (1) The tWW (G25:U20 in reverse wobble) base pairing may result in a preferred conformation so that C75:N<sub>3</sub> is in close proximity to G1:O<sub>1P</sub> and G2:O<sub>1P</sub> in comparison to tWH base pairing. (2) The B-site Mg<sup>2+</sup> binding brings an induced fit, resulting in a favorable tWW conformation. (3) The tWH base pairing may be not able to bring C75:N<sub>3</sub> close to G1:O<sub>1P</sub> and G2:O<sub>1P</sub>, except in the case of Mg<sup>2+</sup> initially placed at the C-site.

Such average values can be used to rationalize the possible positive  $pK_a$  shift of C75:N<sub>3</sub>.

In the tWW\_RT\_C75°M<sub>B</sub> simulation, G25:U20 was initially in a tWW conformation and Mg<sup>2+</sup> was placed at the B-site with an outer-sphere coordination at G25:N<sub>7</sub>. The G25:U20 tWW conformation is stable throughout the simulation (total 250 ns), as indicated by the strong hydrogen bonds between G25:N<sub>1</sub>/U20:O<sub>4</sub> and U20:N<sub>3</sub>/G25:O<sub>6</sub>, G25 remained in a syn conformation ( $\chi = 75.5^\circ$ ), and the Mg<sup>2+</sup> kept its initial binding mode to G25:N<sub>7</sub> with an average distance of 4.90 Å.

In the tWW\_RT\_C75°M<sub>C</sub> simulation, Mg<sup>2+</sup> was initially placed at the C-site where it coordinated with G1:N<sub>1</sub>. The Mg<sup>2+</sup> did not maintain its coordination with G1:N<sub>1</sub> and was eventually (after ~100 ns) ejected from the C-site (average distance 7.41 Å). Interestingly, the G25:U20 tWW conformation, although it was stable for most of the simulation, was weakened, as indicated by the fact that the two key hydrogen bonds, G25:N<sub>1</sub>/U20:O<sub>4</sub> and U20:N<sub>3</sub>/G25:O<sub>6</sub>, were maintained in only 60% of the snapshots (see the criteria in Table 1), down from around 80% in the tWW\_RT\_C75°M<sub>B</sub> simulation. G25 showed strong deviation in its  $\chi$  angle and switched toward an anti conformation after Mg<sup>2+</sup> left the C-site (average  $\chi = 151.4^\circ$ ). The tWW\_RT\_C75°M<sub>C</sub> simulation thereafter essentially became identical with the tWW\_RT\_C75° simulation discussed below.

In the tWW\_RT\_C75° simulation, no Mg<sup>2+</sup> ion was placed at either the B-site or C-site. Although the overall active site integrity is more or less maintained, the G25:U20 tWW conformation is significantly weakened, as indicated by the two key hydrogen bonds (G25:N<sub>1</sub>/U20:O<sub>4</sub> and U20:N<sub>3</sub>/G25:O<sub>6</sub>) being present only 20% of the time. In addition, the loss of these hydrogen-bonding interactions correlates with rotation of G25 toward the anti conformation (average  $\chi = -169.2^\circ$ ). A similar tWW\_RT\_C75° simulation (labeled as tWW\_RT\_C75°\_fix in Table 1) was performed in which two key hydrogen bonds, G25:N<sub>1</sub>/U20:O<sub>4</sub> and U20:N<sub>3</sub>/G25:O<sub>6</sub>, were restrained throughout the simulation. Despite enforcing the tWW pairing, G25 appeared to prefer an anti conformation (average  $\chi = -167.1^\circ$ ). Another independent 300 ns simulation was performed and delivered qualitatively the same results (see Table S1 in the Supporting Information).

The above simulation results of tWW\_RT\_C75° with different initial binding modes of Mg<sup>2+</sup> suggest that the B-site Mg<sup>2+</sup> is critical for maintaining the syn glycosidic torsion and hence the tWW (reverse wobble) conformation, likely through water-mediated coordination at G25:N<sub>7</sub>. The G25 base prefers an anti conformation even when a tWW base pair is enforced (tWW\_RT\_C75°\_fix). This is consistent with crystallographic evidence<sup>44</sup> and chemical probing experiments.<sup>45</sup>

Another set of simulations were performed with G25:U20 initially in the tWH conformation, again with various initial binding modes of Mg<sup>2+</sup>. In all simulations the tWH conformation was not stable, as indicated by the necessary hydrogen bonds weakening until essentially no base pair interaction was present. The tWH conformation is therefore not stable in the reactant state, regardless of the Mg<sup>2+</sup> binding mode. This observation is consistent with the fact that the G25 position was not resolved and is likely flexible in the crystal without Mg<sup>2+</sup> ions.<sup>42</sup> When Mg<sup>2+</sup> was initially placed at the B-site (tWH\_RT\_C75°M<sub>B</sub>), it quickly coordinated directly with G1:O<sub>2P</sub> (average distance 1.87 Å). When Mg<sup>2+</sup> was initially placed at the C-site (tWH\_RT\_C75°M<sub>C</sub>), it maintained its inner sphere coordination with G1:N<sub>1</sub> for the first

100 ns and then moved to outer-sphere coordination (average distance 5.62 Å). In all tWH simulations, G25 kept an anti conformation (average  $\chi$  between  $-50^\circ$  and  $-80^\circ$ ).

The  $\langle(1/r_1) + (1/r_2)\rangle$  term in Table 1 can be treated as a measure of the electrostatic effect on C75:N<sub>3</sub> from G1:O<sub>1P</sub> and G2:O<sub>1P</sub>. Here, in all simulations of the reactant state in the tWW conformation, these values are almost identical, indicating that in the tWW conformation C75 positions itself independently of the Mg<sup>2+</sup> binding mode. The only exception is when Mg<sup>2+</sup> is absent, in which case the value is slightly higher and G1:O<sub>1P</sub>/G2:O<sub>1P</sub> are enforced to be in a tWW conformation (tWW\_RT\_C75°\_fix,  $\langle(1/r_1) + (1/r_2)\rangle = 0.46$ ). Nevertheless, in tWH simulations the  $\langle(1/r_1) + (1/r_2)\rangle$  value becomes much smaller when the Mg<sup>2+</sup> is initially placed at the B-site (tWH\_RT\_C75°M<sub>B</sub>,  $\langle(1/r_1) + (1/r_2)\rangle = 0.20$ ) or when Mg<sup>2+</sup> is absent from both sites (tWH\_RT\_C75°,  $\langle(1/r_1) + (1/r_2)\rangle = 0.31$ ). Although the value is much higher when Mg<sup>2+</sup> is initially placed at the C-site (tWH\_RT\_C75°M<sub>C</sub>,  $\langle(1/r_1) + (1/r_2)\rangle = 0.49$ ), this could be a simulation artifact, as this Mg<sup>2+</sup> gradually dissociates from the C-site and the system is likely to eventually behave in the same way as the tWH\_RT\_C75° case after the Mg<sup>2+</sup> is no longer bound.

All of the simulation results of the reactant state mentioned here imply the following (a) The tWH conformation is unstable in the reactant state, regardless of the Mg<sup>2+</sup> binding mode. (b) The tWW (reverse wobble) pair is stable in the reactant state with C75:N<sub>3</sub> in close proximity to G1:O<sub>1P</sub> and G2:O<sub>1P</sub>; this juxtaposition is apparently induced by Mg<sup>2+</sup> binding at the B-site. (c) The tWH base pairing may not be able to position C75:N<sub>3</sub> close to G1:O<sub>1P</sub> and G2:O<sub>1P</sub>.

A possible mechanistic pathway that draws together these simulations, shown in Figure 3, is as follows. In the reactant state, a tWH pair is more stable than a tWW pair if there is no Mg<sup>2+</sup> bound near the active site. With Mg<sup>2+</sup> at the B-site, a tWW pair is preferred and leads to a repositioning of C75:N<sub>3</sub> such that it is proximal to G1:O<sub>1P</sub> and G2:O<sub>1P</sub>, thereby increasing its  $pK_a$ . Note that here it is the conformation that leads to the  $pK_a$  shift, not the Mg<sup>2+</sup> ion. In fact, in the tWW conformation the B-site Mg<sup>2+</sup> will likely shift the  $pK_a$  of C75 downward, not upward, due to Coulombic repulsion.

Hence, the B-site Mg<sup>2+</sup> plays a structural role resulting in a conformation favorable for the  $pK_a$  shift of C75. This scenario is consistent with the second reaction channel model proposed by Nakano et al.,<sup>90</sup> in which the contribution of a structural metal ion was deduced from kinetic experiments. Moreover, this model explains the discrepancy that C75 is observed without a significant  $pK_a$  shift using NMR<sup>37</sup> but shows a ~2.0 unit shift using Raman crystallography, as well as a slight anti correlation with the Mg<sup>2+</sup> concentration.<sup>35</sup> The HDVr could be trapped in the tWW conformation in the Raman crystallography experiments but in an equilibrium between tWW/tWH conformations under solution conditions.

**Protonation of C75 Leads to Stabilization of the tWH/cWW Conformations That Favor Binding of a Mg<sup>2+</sup> at the C-Site, Where the Mg<sup>2+</sup> Ion May Act as the Base Catalyst Through a Coordinating Water.** There is abundant experimental and theoretical evidence that protonation of C75 is a prerequisite for the HDVr reaction. Nevertheless, this change in the C75 charge state will necessarily cause a nontrivial electrostatic perturbation and the active site environment will respond accordingly. Hence, it is necessary to identify possible conformational changes and Mg<sup>2+</sup> binding modes that are sensitive to the protonated C75. Similarly to Table 1,

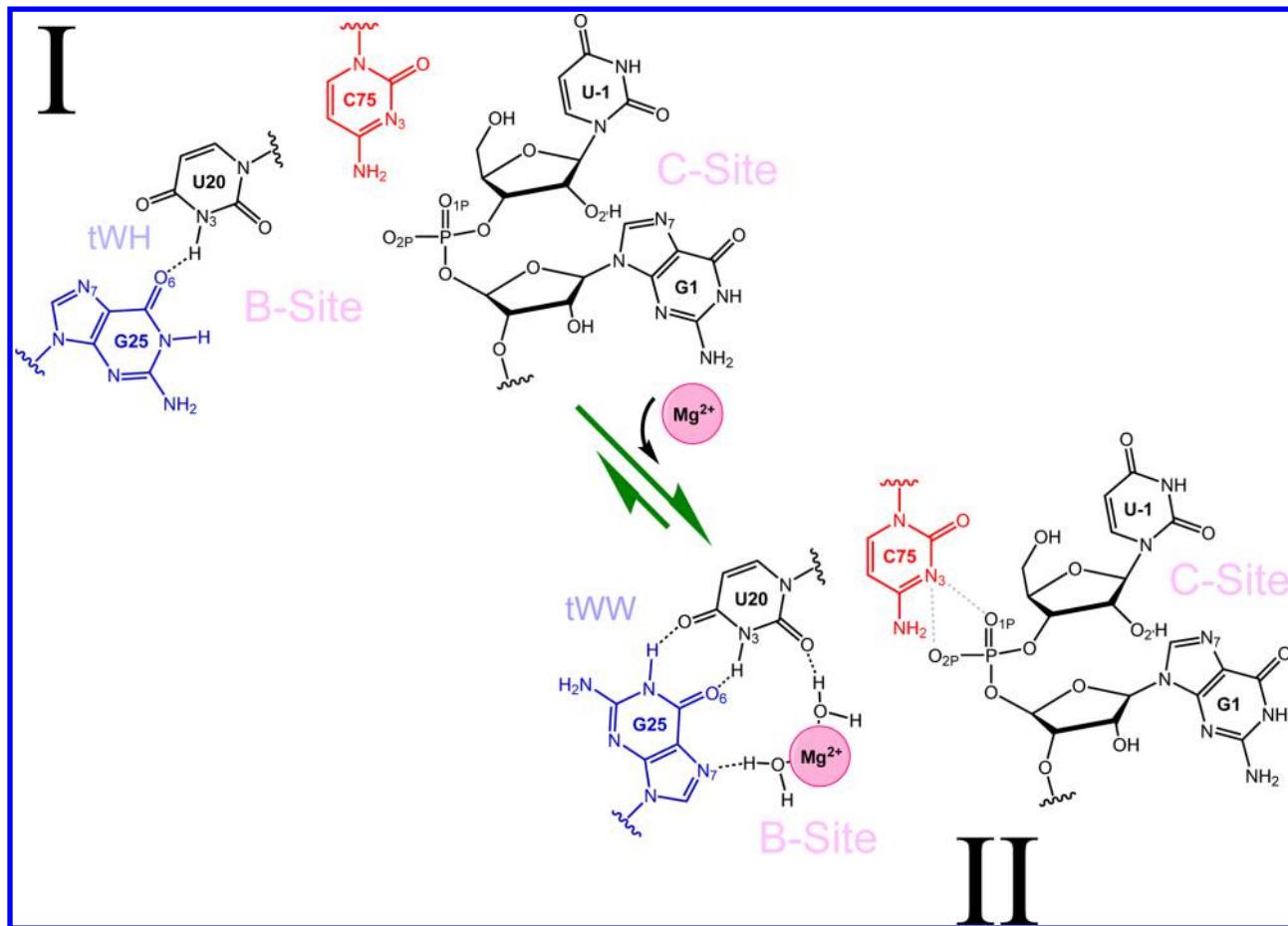


Figure 3. Proposed dynamic model in the reactant state. The B-site  $Mg^{2+}$  binding leads to the induced fit of the tWW conformation of G25:U20.

**Table 2. Key Geometric Parameters for the C75 Protonated Reactant State Simulations (RT\_C75<sup>+</sup>)<sup>a</sup>**

simulation	hydrogen bond			G25: $\chi$	distance to $Mg^{2+}(B)$				distance to $Mg^{2+}(C)$	
	G25:N <sub>1</sub> /U20:O <sub>4</sub>	U20:N <sub>3</sub> /G25:O <sub>6</sub>	U20:O <sub>2</sub> /G25:N <sub>1</sub>		U-1:O <sub>2'</sub>	G1:O <sub>2P</sub>	G25:N <sub>7</sub>	U20:O <sub>2</sub>	U-1:O <sub>2'</sub>	G1:N <sub>7</sub>
tWW_RT_C75 <sup>+</sup> M <sub>B</sub>	0.3	15.0	8.4	65.5(34.3)	7.83(0.28)	4.04(0.26)	6.03(1.31)	5.39(0.93)		
tWW_RT_C75 <sup>+</sup> M <sub>C</sub>	9.3	6.5	28.2	120.7(61.9)					4.09(0.26)	2.23(0.11)
tWW_RT_C75 <sup>+</sup> M <sub>C_fix</sub>	81.1	86.4		64.5(11.7)					8.14(1.70)	2.20(0.08)
tWW_RT_C75 <sup>+</sup>	83.1	81.4		63.9(10.3)						
tWH_RT_C75 <sup>+</sup> M <sub>B</sub>				-65.9(17.0)	7.99(0.21)	4.31(0.26)	7.43(1.05)	4.17(0.36)		
tWH_RT_C75 <sup>+</sup> M <sub>C</sub>				85.9	39.4	-62.8(14.8)			5.67(1.12)	2.26(0.35)
tWH_RT_C75 <sup>+</sup>				73.0	31.6	-80.4(16.8)				

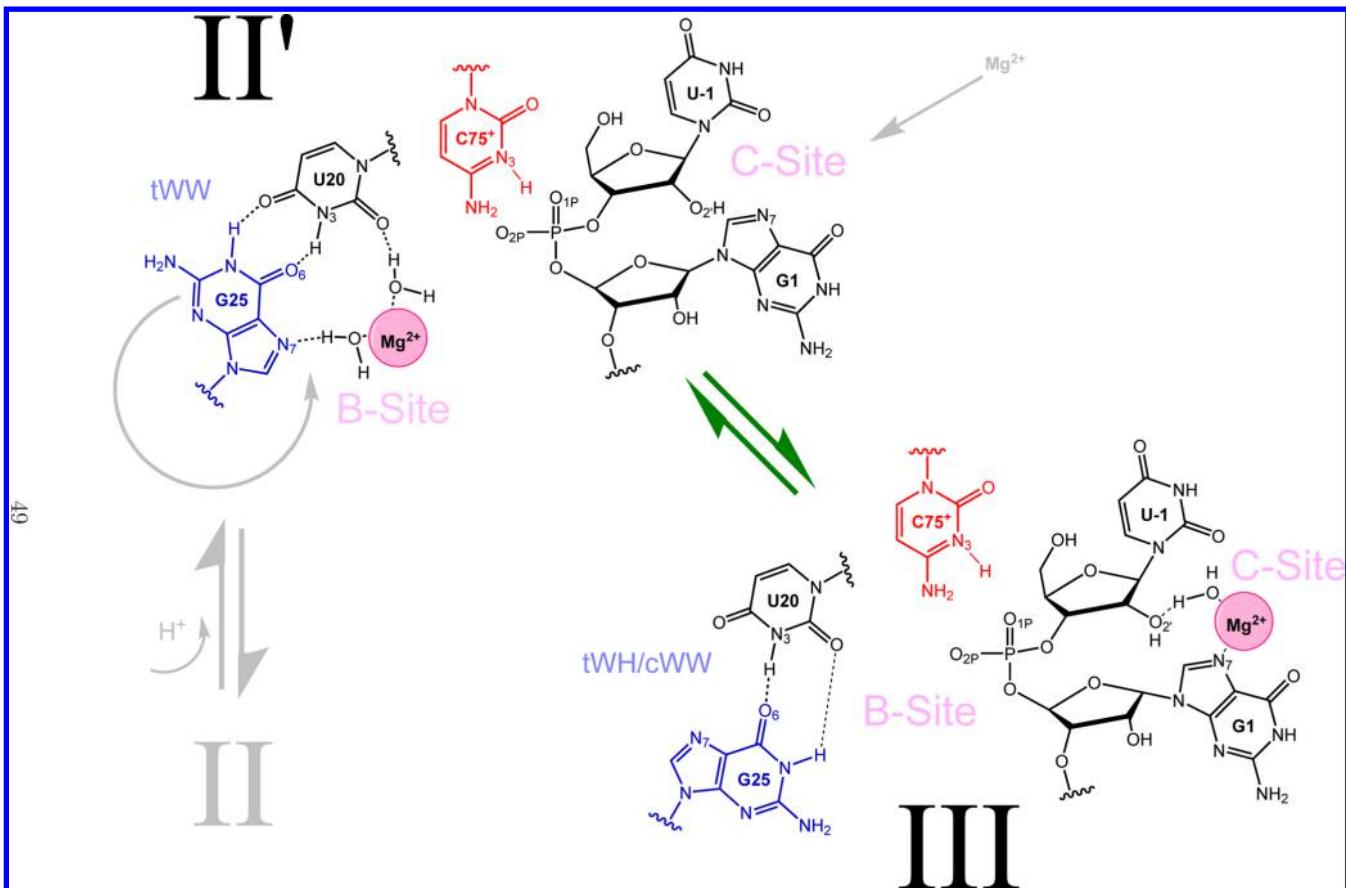
<sup>a</sup>Notations used: Initial G25:U20 base pairing, tWW (reverse wobble) and tWH (Figure 2); M<sub>B</sub>, Mg<sup>2+</sup> initially placed at the B-site (near G25:N<sub>7</sub>); M<sub>C</sub>, Mg<sup>2+</sup> initially placed at the C-site (near G1:N<sub>7</sub>); C75<sup>+</sup>, C75 is protonated and positively charged; C75°, C75 is neutral; RT, reactant state. Entries are calculated from sampling of the range 50–300 ns from the 300 ns trajectory for each simulation condition with a sampling free frequency of 100 ps, resulting in 2500 data points for every simulation. Distances are given in Å, and hydrogen bond occupations (given as percentages) are defined as the percentage of the snapshots in which the distance between the proton donor and the proton acceptor  $r \leq 3.5$  Å and the H-bond angle  $\theta \geq 150^\circ$ . The numbers are average values, and corresponding standard deviations are given in parentheses. Simulation results suggest that when C75 is protonated and the Mg<sup>2+</sup> is present (at either the B-site or the C-site), the G25:U20 tWW conformation is not stable. The preferred G25:U20 tWH/cWW conformation with Mg<sup>2+</sup> at the C-site suggests a likely role of this Mg<sup>2+</sup> ion is to act as the base catalyst through a hydroxide.

Table 2 gives key geometric parameters characterizing the base pairing of G25:U20 and the Mg<sup>2+</sup> binding of the B- and C-sites, but now in the presence of protonated C75.

In the tWW\_RT\_C75<sup>+</sup>M<sub>B</sub> simulation, Mg<sup>2+</sup> remains at the B-site, but the tWW conformation is clearly weakened, although G25 retains its syn conformation (average  $\chi = 65.5^\circ$ ). It is clear that, once C75 is protonated, the B-site Mg<sup>2+</sup> no longer prefers coordination with G25:N<sub>7</sub> (average distance 6.03 Å) and during the simulation it never gets close enough to

the nucleophile (U-1:O<sub>2'</sub>) to possibly act as a base catalyst (average distance 7.83 Å).

The two characteristic hydrogen bonds of the tWW conformations are essentially lost in the tWW\_RT\_C75<sup>+</sup>M<sub>B</sub> simulation. Hence the tWW form is not stable after C75 is protonated. This apparently results from electrostatic repulsion between the B-site Mg<sup>2+</sup> and C75<sup>+</sup>, which pushes the B-site Mg<sup>2+</sup> away from the active site (II' in Figure 4). Thus, the simulation results so far suggest that the B-site Mg<sup>2+</sup> creates an



**Figure 4.** Proposed dynamic model with protonated C75<sup>+</sup> (II') and the resulting conformation after the B-site Mg<sup>2+</sup> leaves, the G25:U20 switches from tWW to tWH/cWW, and another Mg<sup>2+</sup> enters the C-site (III).

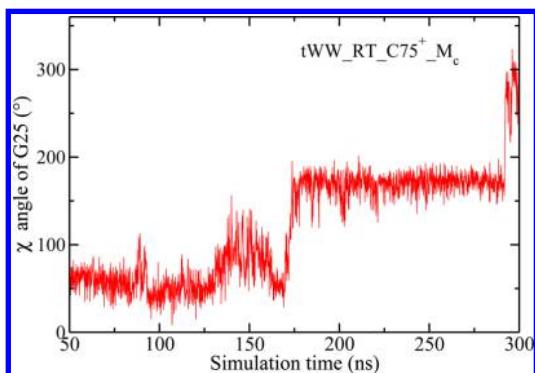
environment that increases the probability of C75 protonation; i.e., increases its pK<sub>a</sub>. However, once C75 is protonated the metal ion loses its binding to G25:N<sub>7</sub>, which favors dissociation from the active site (see below).

In the tWW\_RT\_C75<sup>+</sup>M<sub>C</sub> simulation, Mg<sup>2+</sup> remains at the C-site and again the tWW hydrogen bonds are lost. However, now G25 rotates toward an anti conformation (average  $\chi = 120.7^\circ$ ) after 170 ns (switching from  $\sim -50^\circ$  to  $\sim 180^\circ$  and then to  $\sim 275^\circ$ , Figure 5) and the tWW conformation is lost,

potentially on its way to reach a tWH or cWW geometry. The Mg<sup>2+</sup> steadily coordinates to the nucleophile through a water molecule and is poised to act as the base catalyst (average distance 4.09 Å).

We further investigated the relationship between C-site Mg<sup>2+</sup> binding and the G25:U20 tWW conformation in an additional tWW\_RT\_C75<sup>+</sup>M<sub>C</sub> simulation (denoted as tWW\_RT\_C75<sup>+</sup>M<sub>C</sub>\_fix), in which harmonic biasing potentials were applied to enforce the hydrogen bonds between the bases. The results, given in Table 2, show that the C-site Mg<sup>2+</sup> still remains at the C-site during the simulation (average distance 2.23 Å) but no longer coordinates to the nucleophile, U-1:O<sub>2'</sub> (average distance 8.14 Å). The observation that the B-site Mg<sup>2+</sup> cannot stay with the tWW conformation was verified by another independent 300 ns tWW\_RT\_C75<sup>+</sup>M<sub>2</sub> simulation, where two Mg<sup>2+</sup> ions were added at both B- and C-sites initially (Table S2 in the Supporting Information).

After closely examining the conformational differences between tWW\_RT\_C75<sup>+</sup>M<sub>C</sub> and tWW\_RT\_C75<sup>+</sup>M<sub>C</sub>\_fix, we find that when G25:U20 is enforced in a tWW (reverse wobble) conformation, G25:N<sub>7</sub> favors the binding of sodium ions in the active site to form a relatively rigid network of interactions with nearby residues. Conversely, when a Mg<sup>2+</sup> is present at the C-site, the binding of sodium ions in the active site will be unfavorable due to simple electrostatic repulsion. The result is that, with C-site Mg<sup>2+</sup> binding, the conformational preference of G25:U20 is shifted from tWW to tWH/cWW. This observation is also supported by the tWW\_RT\_C75<sup>+</sup> simulation where no Mg<sup>2+</sup> is in either site: the tWW



**Figure 5.** Time series of the glycosidic bond torsion (the  $\chi$  angle, defined as the torsion between O<sub>4'</sub>-C<sub>1'</sub>-N<sub>9</sub>-C<sub>4</sub>) of G25 for the tWW\_RT\_C75<sup>+</sup>M<sub>C</sub> simulation. The G25: $\chi$  angle changes from  $\sim 50^\circ$  (a syn conformation), to  $\sim 180^\circ$  after 175 ns, and then to over  $250^\circ$  (a clear anti conformation). In order to demonstrate the continuous changes, the  $\chi$  angles shown here are in the range between 0 and 360°, while the range of  $-180$  to  $180$ ° is used in other places.

conformation is kept during the whole simulation time (300 ns), as strongly as in the simulations with biasing potentials (tWW\_RT\_C75<sup>+</sup>M<sub>C</sub>\_fix).

Another set of simulations were performed with C75 protonated but G25:U20 initially in its tWH conformation and Mg<sup>2+</sup> alternately placed at the B-site or C-site or Mg<sup>2+</sup> absent altogether. The results suggest (Table 2) that binding of the B-site Mg<sup>2+</sup> would destabilize the G25:U20 base interaction as all hydrogen bonds are lost, which is further verified by a repeated simulation (Table S2 in the Supporting Information). Nevertheless, the tWH conformation is somehow kept with some distribution of the cWW conformation in both simulations with C-site Mg<sup>2+</sup> or no Mg<sup>2+</sup>. The Mg<sup>2+</sup> in tWH\_RT\_C75<sup>+</sup>M<sub>C</sub> also forms water-mediated coordination to the nucleophile (U-1:O<sub>2'</sub>) to be ready to act as the base catalyst during most of the simulation time (average distance 5.67 Å), although not as strongly as in the case of tWW\_RT\_C75<sup>+</sup>M<sub>C</sub>. Note that, in tWW\_RT\_C75<sup>+</sup>M<sub>C</sub>, the tWW conformation is not stable and seems to move toward tWH/cWW conformations.

As graphically summarized in Figure 4, we conclude the following from the results in Table 2: after C75 is protonated, the interaction of the B-site Mg<sup>2+</sup> with G25 is weakened and may favor dissociation. Without the B-site Mg<sup>2+</sup> it is unfavorable for the G25:N<sub>7</sub> binding pocket to face the active site and G25:U20 prefers the tWH/cWW conformation. The resulting rearrangement of the active site creates a large space at the C-site, resulting in recruitment of another Mg<sup>2+</sup> into the C-site. This new Mg<sup>2+</sup> is in a position to serve as the base catalyst (Table 2 and III in Figure 4). This model is consistent with the third channel of HDVr reaction suggested by Nakano et al.<sup>90</sup> and Cerrone-Szakal et al.,<sup>36</sup> which involves both structural and catalytic Mg<sup>2+</sup> ions.

**Deprotonation of U-1:O<sub>2'</sub> Causes Rotation of U-1, Leading to an Active, Inline Conformation.** After the nucleophile (U-1:O<sub>2'</sub>) is deprotonated, a necessary condition for the reaction to proceed is the formation of a conformation whereby the nucleophile is poised to make an inline attack to the scissile phosphate: e.g., the U-1:O<sub>2'</sub>–G1:P–G1:O<sub>S'</sub> angle of ~160–180°. Such a conformation has not yet been directly observed from the electron density in any reported crystal structure of the HDVr, although a model structure was recently proposed on the basis of the Hammerhead ribozyme active site structure.<sup>44</sup> In all known structures in which U-1 is resolved (mainly C75U mutants<sup>42</sup>), a large-scale rotation of U-1 would be needed to reach an inline conformation. In simulations departing from one such structure (activated by computational mutagenesis), we recently reported the observation of the spontaneous rotation of U-1 resulting in an inline conformation.<sup>59</sup>

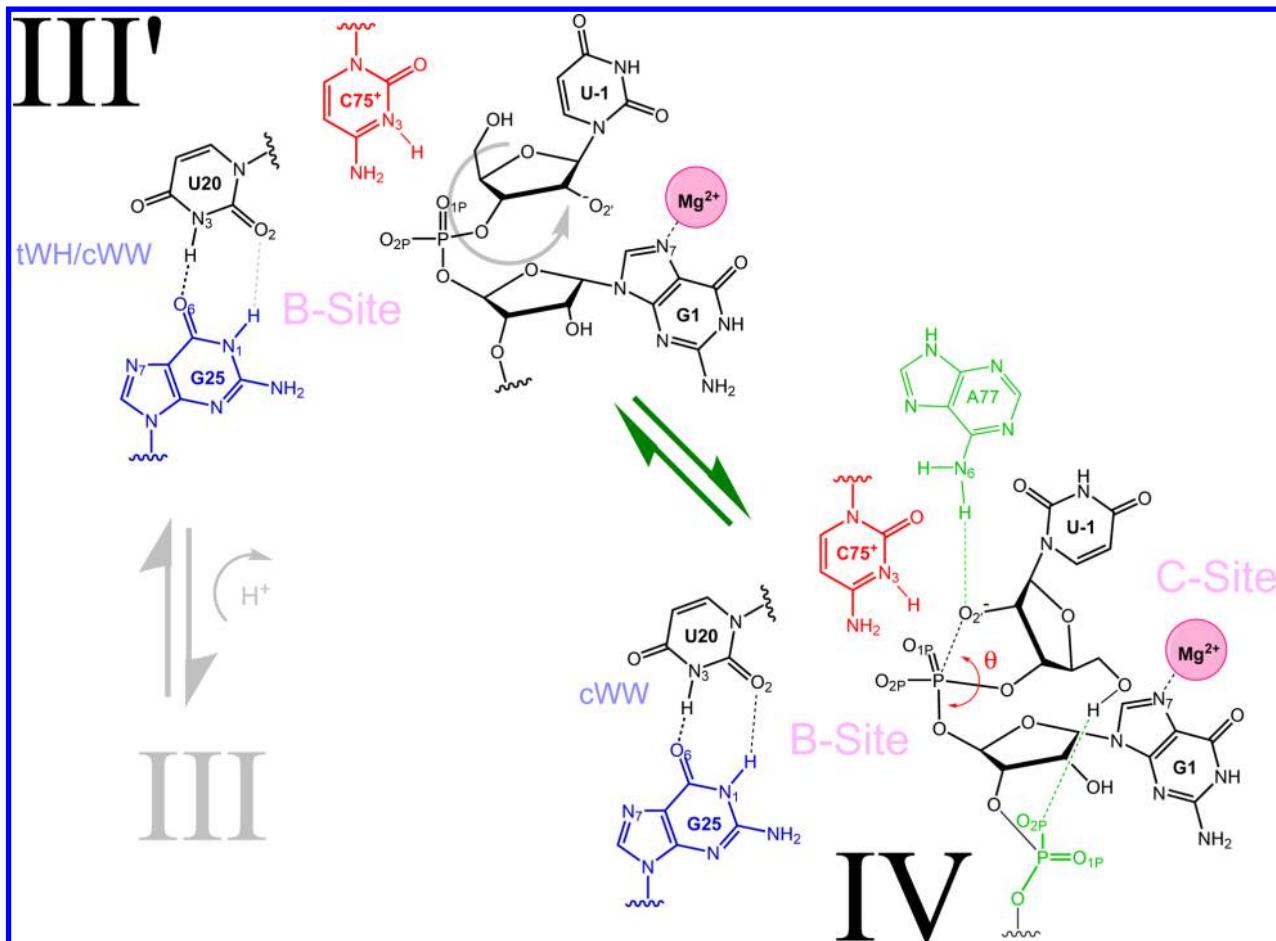
Table 3 gives the results of simulations with U-1:O<sub>2'</sub> in a deprotonated state (denoted as dRT) with different G25:U20 conformations and Mg<sup>2+</sup> binding modes. In the tWW\_dRT\_C75<sup>+</sup>M<sub>B</sub> simulations, the Mg<sup>2+</sup> remains bound and retains water-mediated contacts with G25:N<sub>7</sub> (average distance 4.72 Å) and G1:O<sub>2P</sub> (average distance 4.28 Å). However, the tWW conformation is weakened, as indicated by the G25:N<sub>1</sub>/U20:O<sub>4</sub> hydrogen bond percentage dropping to 33.2%. The average inline angle was 95.7°, and the inline angle was never larger than 120° during the 300 ns simulation time.

In the tWW\_dRT\_C75<sup>+</sup>M<sub>C</sub> simulation, the Mg<sup>2+</sup> again remained in its initial C-site binding mode (average distance to G1:N<sub>7</sub> is 2.19 Å), but with an average inline angle of 141.7°. In 96.0% of the time during simulation the inline angle was again

**Table 3. Key Geometric Parameters in the Activated Precursor State Simulations (dRT\_C75<sup>+</sup>)<sup>a</sup>**

	hydrogen bond						distance to C75:N <sub>3</sub>			inline angle			distance to Mg <sup>2+</sup> (B)			
	G25:N <sub>1</sub> /U20:O <sub>4</sub>	U20:N <sub>3</sub> /G25:O <sub>6</sub>	U20:O <sub>3</sub> /G25:N <sub>1</sub>	G25:X	G1:O <sub>1P</sub>	G1:O <sub>2P</sub>	G1:O <sub>S'</sub>	av	% (>120°)	U-1:O <sub>2'</sub>	G1:O <sub>2P</sub>	G25:N <sub>7</sub>	U20:O <sub>2</sub>	G1:O <sub>2</sub>	U-1:O <sub>2'</sub>	distance to Mg <sup>2+</sup> (C)
tWW_dRT_C75 <sup>+</sup> M <sub>B</sub>	33.2	81.9	0.2	70.5(15.3)	2.90(0.25)	3.71(0.47)	5.06(0.26)	95.7(6.2)	141.7(11.1)	96.0	6.19(0.95)	4.28(0.36)	4.72(0.73)	4.80(0.81)	7.72(0.76)	2.19(0.09)
tWW_dRT_C75 <sup>+</sup> M <sub>C</sub>	6.9	89.0	65.8(11.1)	8.38(0.56)	7.83(0.53)	6.78(0.66)	—	—	—	0.0	9.50(0.82)	6.43(1.48)	8.44(1.67)	6.58(1.25)	7.19(0.67)	2.21(0.10)
tWH_dRT_C75 <sup>+</sup> M <sub>B</sub>	0.0	—	—	-66.5(18.3)	4.17(0.53)	3.31(0.26)	5.29(0.24)	96.5(6.0)	—	—	—	—	—	—	—	—
tWH_dRT_C75 <sup>+</sup> M <sub>C</sub>	84.1	72.9	-84.7(10.3)	3.71(0.79)	3.23(0.44)	5.12(0.30)	129.1(18.0)	78.3	—	—	—	—	—	—	—	—
tWH_dRT_C75 <sup>+</sup>	0.7	—	-44.5(10.3)	4.88(1.67)	3.76(0.72)	5.74(0.82)	94.0(8.8)	0.2	—	—	—	—	—	—	—	—

<sup>a</sup>Notations used: initial G25:U20 base pairing, tWW (reverse wobble) and tWH (Figure 2); M<sub>B</sub>, Mg<sup>2+</sup> initially placed at the B-site (near G25:N<sub>7</sub>); M<sub>C</sub>, Mg<sup>2+</sup> initially placed at the C-site (near G1:N<sub>7</sub>); C75<sup>+</sup>, C75 is protonated and positively charged; C75°, C75 is neutral; dRT, activated reactant state (the nucleophile (U-1:O<sub>2'</sub>) is deprotonated). Entries are calculated from sampling of the range 50–300 ns from the 300 ns trajectory for each simulation condition with a sampling free frequency of 100 ps, resulting in 2500 data points for every simulation. Distances are given in Å, and hydrogen bond occupations (given as percentages) are defined as the percentage of the snapshots in which the distance between the proton donor and the proton acceptor *r* ≤ 3.5 Å and the H-bond angle *θ* ≥ 150°. The numbers are average values, and corresponding standard deviations are given in parentheses. Simulation results suggest the following (1) The initial tWH conformation with Mg<sup>2+</sup> placed at the C-site is the most likely combination where the simultaneous U-1 rotation will occur after the nucleophile (U-1:O<sub>2'</sub>) is deprotonated (Figure 6). (2) The resulting strong cWW conformation hydrogen binding (U20:N<sub>3</sub>/G25:O<sub>6</sub> and U20:O<sub>2</sub>/G25:N<sub>7</sub>, Figure 2) may be important for the U-1 rotation, as well as two possible anchor functional groups (see text), A77:N<sub>6</sub> and G2:O<sub>2P</sub>. (3) The B-site Mg<sup>2+</sup> may in fact inhibit such a rotation, as there is no population of inline angle greater than 120° when Mg<sup>2+</sup> is at a B-site regardless of the initial G25:U20 conformation.



**Figure 6.** Proposed dynamic model of the activated precursor state, where the nucleophile, U-1:O<sub>2'</sub>, is deprotonated (III') and the resulting conformation after U-1 rotates to reach an inline conformation ready for the next step of the reaction, the nucleophilic (S<sub>N</sub>2-like) reaction (IV). The G25:U20 now forms a stronger cWW pairing.

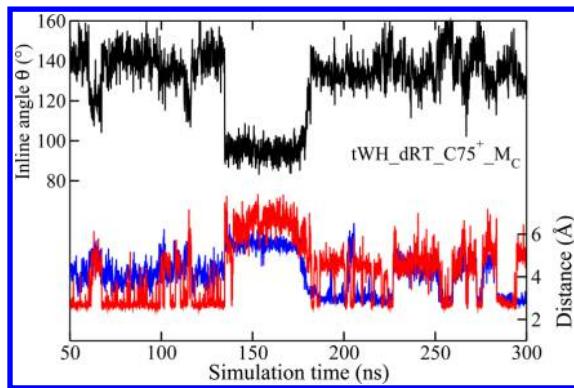
significantly weakened. It is worth noting that, although the tWW conformation was weakened, the U20:N<sub>3</sub>/G25:O<sub>6</sub> hydrogen bond is still strongly held 89.0% of the time. After closely examining the structures in the tWW\_dRT\_C75<sup>+</sup>M<sub>C</sub>, although the inline angle is apparently sufficient for reaction to occur, we found that the U-1 rotation brings U-1:O<sub>O2'</sub> closer to the phosphate group of G2 and in fact results in a conformation where C75 is far from ideal as a general acid, as the average distances between C75:N<sub>3</sub> and G1:O<sub>1P</sub> (8.38 Å), G1:O<sub>2P</sub> (7.83 Å), and G1:O<sub>5'</sub> (6.78 Å) all increase significantly. Hence, a tWW conformation with a C-site bound Mg<sup>2+</sup> is unlikely to be the most active conformation, even though a proper inline angle was obtained.

With G25:U20 initially in the tWH conformation (tWH\_dRT\_C75<sup>+</sup>M<sub>B</sub>), the B-site Mg<sup>2+</sup> appears to be less stable and the base pairing is disrupted. All key hydrogen bonds are broken and no inline conformations are observed. On the other hand, when the Mg<sup>2+</sup> is initially placed at the C-site (tWH\_dRT\_C75<sup>+</sup>M<sub>C</sub>), the Mg<sup>2+</sup> remains bound during the simulation (average distance to G1:N<sub>7</sub> is 2.21 Å) and the tWH conformation transitions to a strong cWW conformation, producing a significant population of inline conformations (78.3% of inline angle population >120°). C75:N<sub>3</sub> is also in a plausible position to act as the general acid. The simulation was repeated to further verify the observations (Table S3 in the Supporting Information). Upon closer examination of the

trajectory, two interactions (possibly water mediated) could be seen as the driving force for U-1 rotation. Figure 7 shows the time series of the inline angle along with two distances between U-1 and G1/A77 (Figure 2). These distances are useful markers for interactions that are only present in active, inline conformations. The results are consistent with the detrimental effects of A77 mutants on HDV ribozyme function<sup>91</sup> as well as observation of phosphorothioate interference at G2:O<sub>2P</sub>,<sup>76</sup> for which, to our knowledge, no structural rationale has previously been suggested. Note that the present simulation work does not provide into the insights of the time scale or the free energy barrier of U-1 rotation.

In summary, Table 3 suggests the following. (a) The initial tWH conformation with Mg<sup>2+</sup> placed at the C-site is the most likely state from which U-1 rotation will occur after the nucleophile is deprotonated (Figure 6). (b) The resulting strong cWW conformation hydrogen bonding (84.1% for U20:N<sub>3</sub>/G25:O<sub>6</sub> and 72.9% for U20:O<sub>2'</sub>/G25:N<sub>1</sub>, Figure 2), as well as two possible anchor points, A77:N<sub>6</sub> and G2:O<sub>2P</sub>, may be important for the U-1 rotation. (c) The B-site Mg<sup>2+</sup> may in fact inhibit such a rotation, as there is no population of inline angle greater than 120° when a Mg<sup>2+</sup> is at the B-site, regardless of the initial G25:U20 conformation.

**Movement of the Protonated C75<sup>+</sup> to a Position Poised for General Acid Catalysis and Ejection of the C-Site Mg<sup>2+</sup> as the Reaction Proceeds.** Following (or concurrent with)



**Figure 7.** Time series of the inline attacking angle ( $\theta(\text{U-1:O}_2\text{-G1:P-G1:O}_5)$ , in black) and two possible hydrogen bond pairs holding the inline conformation,  $\text{U-1:O}_2/\text{A77:N}_6$  (blue) and  $\text{G1:O}_{2P}/\text{U-1:O}_5'$  (red), in the simulation  $\text{tWH\_dRT\_C75}^+\text{M}_C$  where a simultaneous rotation of U-1 resulting in the inline conformation is observed.

phosphoryl transfer, the proton transfer must occur in order to stabilize the  $\text{G1:O}_5'$  oxide. A wealth of experimental evidence<sup>30,33,35,36</sup> suggests that this proton originates from C75, acting as a general acid, though it has been suggested that C75 becomes positioned for this role only after the precursor state has been reached.<sup>39</sup> Table 4 gives results for simulations exploring various G25:U20 conformations and  $\text{Mg}^{2+}$  binding modes in transition state mimics.

In the  $\text{tWW\_ETS\_C75}^+\text{M}_B$  simulation, where G25:U20 is in a tWW (reverse wobble) conformation and  $\text{Mg}^{2+}$  initially placed at the B-site (near G25:N<sub>7</sub>), the tWW conformation remains stable (~70–80% of the time during the simulation the key hydrogen bonds are kept) and  $\text{Mg}^{2+}$  stays at the B-site. However, the repulsive electrostatic interaction with the B-site  $\text{Mg}^{2+}$  prevents C75<sup>+</sup> from approaching  $\text{G1:O}_5'$  and hence would interfere with its role as general acid (average distance of C75:N<sub>3</sub> to  $\text{G1:O}_5'$  is 7.56 Å). Apparently, in this model the presence of  $\text{Mg}^{2+}$  at the B-site prevents the reaction from proceeding by blocking the movement of C75<sup>+</sup>.

In the  $\text{tWW\_ETS\_C75}^+\text{M}_C$  simulation, where G25:U20 is in a tWW conformation and  $\text{Mg}^{2+}$  is initially placed at the C-site, the tWW conformation is kept and C75:N<sub>3</sub> is closer to  $\text{G1:O}_5'$  but is not yet in the position to act as a general acid (average distance 4.77 Å). This is possibly due to the repulsive electrostatic interaction from sodium ions in the active site recruited by G25:N<sub>7</sub>. A similar situation is observed in the simulation without B-site or C-site  $\text{Mg}^{2+}$  ( $\text{tWW\_ETS\_C75}^+$ ). In addition, the movement of C75<sup>+</sup> to the reaction center causes loss of C-site-bound  $\text{Mg}^{2+}$  (average distance to G1:N<sub>7</sub> jumps to 6.69 Å).

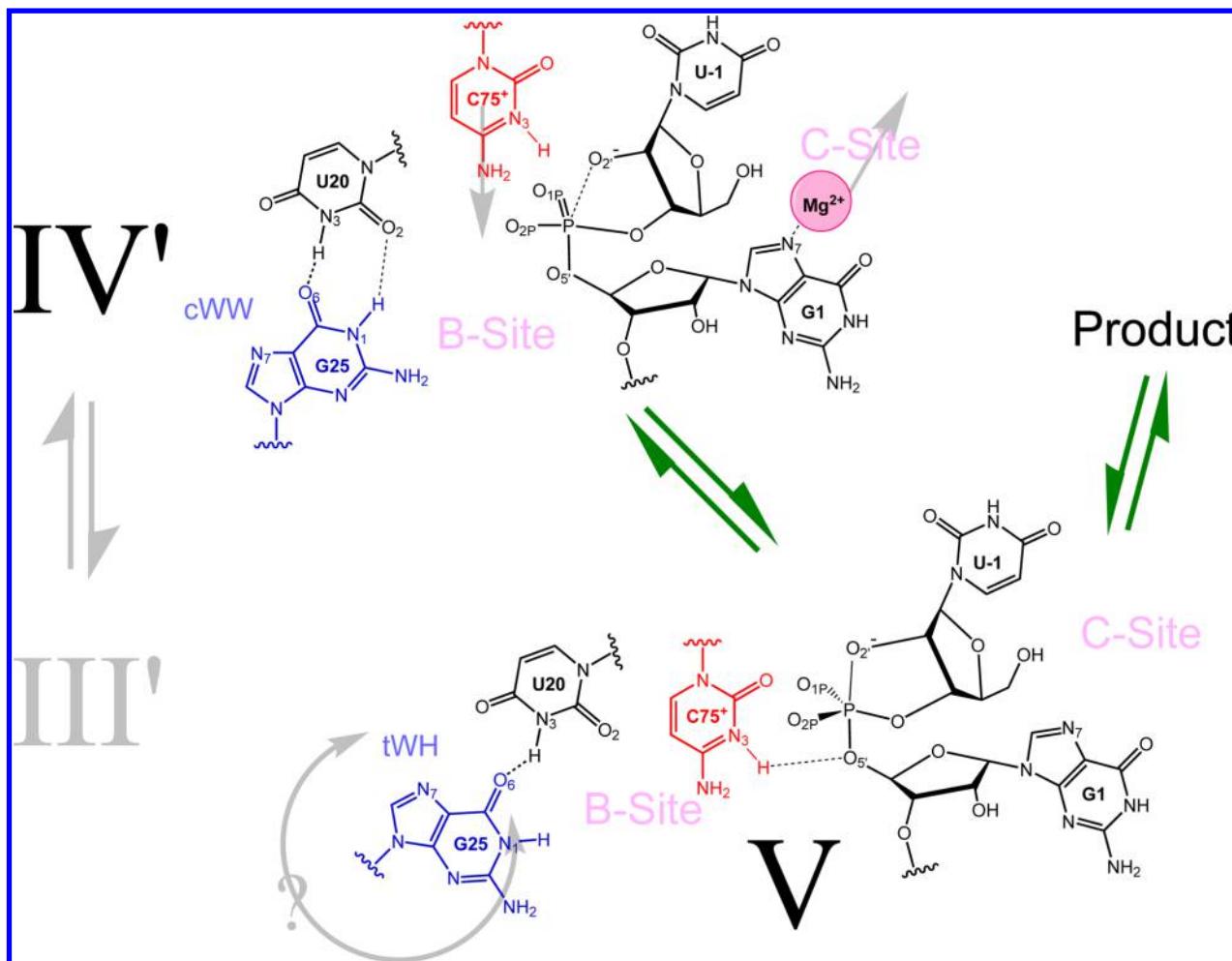
When G25:U20 is initially in its tWH conformation ( $\text{tWH\_ETS\_C75}^+\text{M}_C$ ), G25:U20 forms a strong cWW pairing and C75<sup>+</sup> moves close to  $\text{G1:O}_5'$ , ready to act as the general acid (average distance 3.01 Å) and, at the same time, pushes the  $\text{Mg}^{2+}$  out of the C-site (average distance to G1:N<sub>7</sub> is 7.34 Å). Similar results are observed in the corresponding late transition state mimic simulation ( $\text{tWH\_LTS\_C75}^+\text{M}_C$ ), except that the strong cWW pairing is much weaker.

Hence, the simulation results suggest that  $\text{Mg}^{2+}$  is no longer needed after the S<sub>N</sub>2-like reaction step. The B-site  $\text{Mg}^{2+}$ , as well as sodium ions recruited by the tWW G25:U20 pair, will in fact prevent C75<sup>+</sup> from reaching a position to be the general acid. Therefore, a B-site  $\text{Mg}^{2+}$  is inhibitory for this stage of the reaction. With the G25:U20 pair initially in a tWH conformation (later

**Table 4. Key Geometric Parameters in Transition State Mimic and Product State Simulations (ETS\_C75<sup>+</sup>, LTS\_C75<sup>+</sup>, and ETS\_C75°)<sup>a</sup>**

simulation	hydrogen bond			distance to $\text{Mg}^{2+}$ (B)				distance to $\text{Mg}^{2+}$ (C)		
	G25:N <sub>3</sub> /U20:O <sub>4</sub>	U20:N <sub>3</sub> /G25:O <sub>6</sub>	U20:O <sub>2</sub> /G25:N <sub>1</sub>	G25:N <sub>3</sub> -G1:O <sub>5'</sub> distance	U-1:O <sub>2</sub>	G1:O <sub>2P</sub>	G25:N <sub>7</sub>	U20:O <sub>2</sub>	U-1:O <sub>2</sub>	G1:N <sub>7</sub>
tWW_ETS_C75 <sup>+</sup> M <sub>B</sub>	69.5	80.0	69.3(12.0)	7.56(0.54)	4.57(0.59)	3.95(0.41)	4.43(0.40)	4.73(0.58)	11.13(5.90)	6.69(5.57)
tWW_ETS_C75 <sup>+</sup> M <sub>C</sub>	79.2	85.2	60.8(10.6)	4.77(0.52)	5.83(0.40)	3.01(0.22)	73.0	83.8(10.2)	11.55(3.27)	7.34(5.31)
tWW_ETS_C75 <sup>+</sup>	80.8	82.0	68.7(12.8)	—	—	—	41.6	23.7	83.0(1.83)	6.25(2.47)
tWH_ETS_C75 <sup>+</sup> M <sub>C</sub>	88.7	67.9	116.9(10.0)	—	—	—	0.0	49.8	5.2	—
tWW_LTS_C75 <sup>+</sup> M <sub>C</sub>	88.7	67.9	116.9(10.0)	—	—	—	—	—	3.12(0.45)	—
tWW_PD_C75 <sup>°</sup> M <sub>C</sub>	0.0	—	—	—	—	—	—	—	—	—
tWH_PD_C75 <sup>°</sup> M <sub>C</sub>	—	—	—	—	—	—	—	—	—	—

<sup>a</sup>Notations used: initial G25:U20 base pairing tWW (reverse wobble) and tWH (Figure 2); M<sub>B</sub>, Mg<sup>2+</sup> initially placed at the B-site (near G25:N<sub>7</sub>); M<sub>C</sub>, Mg<sup>2+</sup> initially placed at the C-site (near G1:N<sub>7</sub>); C75<sup>+</sup>, C75 is protonated and positively charged; C75<sup>°</sup>, C75 is neutral; ETS, early transition state mimic; LTS, late-transition state mimic; PD, product state. Entries are calculated from a sampling of the range 50–300 ns from the 300 ns trajectory for each simulation condition with a sampling free frequency of 100 ps, resulting in 2500 data points for every simulation. Distances are given in Å, and hydrogen bond occupations (given as percentages) are defined as as the percentage of the snapshots in which the distance between the proton donor and the proton acceptor  $r \leq 3.5$  Å and the H-bond angle  $\theta \geq 150^\circ$ . The numbers are average values, and corresponding standard deviations are given in parentheses. Simulation results given above suggest that Mg<sup>2+</sup> is no longer needed after the S<sub>N</sub>2-like reaction step. The B-site Mg<sup>2+</sup>, as well as sodium ions recruited by the tWW G25:U20 pair, will in fact prevent C75<sup>+</sup> from reaching the position to be the general acid. Therefore, a B-site Mg<sup>2+</sup> is inhibitory to this stage of the reaction. With the G25:U20 pair initial in tWH (later changed to cWW), C75<sup>+</sup> moves to the general acid position and push the C-site Mg<sup>2+</sup> away from the active site (Figure 8). In the product simulations, the key hydrogen bonds are kept and suggest that both tWW and tWH conformations are stable, but cWW no longer exists. The Mg<sup>2+</sup>, initially placed at the C-site, quickly leaves and enters the bulk solvent in both cases.



**Figure 8.** Proposed dynamic model of the transition states. As the nucleophilic ( $S_N2$ -like) reaction begins (IV'), C75 $^{+}$ :N<sub>3</sub> moves toward the leaving group, G1:O<sub>5'</sub>, and at the same time pushes the C-site Mg<sup>2+</sup> out of the active site (V). The G25:U20 may stay in a weak tWH or change to a tWW conformation after the reaction is completed.

changing to cWW), C75 $^{+}$  moves to the general acid position and pushes the C-site Mg<sup>2+</sup> away from the active site (Figure 8).

Simulation results for the product state with different G25:U20 conformations are also given in Table 4. In both simulations, the key hydrogen bonds are kept, suggesting that they are stable (tWW and tWH but not cWW anymore). The Mg<sup>2+</sup>, initially placed at the C-site, quickly dissociates and enters the bulk solvent. Table 4 and “V” in Figure 8 suggest that the G25:U20 base pairing is not critical after the reaction reaches its late transition state and beyond. Both tWW and tWH seem stable, although weakened, in the product state. This is consistent with the fact that crystal structures of the product state do not have strong tWW G25:U20 base pairing (PDB: 1CX0 and 1DRZ)<sup>27</sup> and that a product structure with a tWH form has also been reported (PDB: 1VC6).<sup>42</sup> Lèvesque et al.<sup>45</sup> in fact also suggested that the tWW is formed *after the cleavage*.

**Novel Dynamic Conformational Model for HDVr Catalysis.** There have been numerous attempts to rationalize and consolidate the various experimental evidence concerning the HDVr catalytic mechanism. Yet a consistent description has been difficult to reach. The key remaining issues are (1) the conformational effects of C75 protonation states, (2) the catalytic role(s) of Mg<sup>2+</sup> and its binding modes, and (3) the catalytically relevant conformations of the active site, especially the G25:U20 base pair.

Previously, we showed that a dynamic model involving distinct changes in conformation and metal binding is likely to be necessary to properly describe the Hammerhead ribozyme reaction.<sup>92–95</sup> In subsequent work regarding the HDVr, we demonstrated that a dynamic model is able to consolidate the experimental evidence regarding the roles of C75 and the necessary conformational change of U-1. This model started with C75 in a position more consistent with activity as a general base and U-1 in an inactive, out-of-inline orientation and ended with C75 in a position consistent with activity as a general acid and U-1 in an active, inline conformation. The latter change took place after a rotation of the U-1 backbone that was spontaneous after changes in the protonation states of C75:N<sub>3</sub> and U-1:O<sub>2'</sub>.<sup>59</sup>

On the basis of the simulation results, here we present an extended dynamic model of the HDVr reaction. The simulation results suggest that the HDVr reaction likely utilizes dynamic changes in the microenvironment of the active site to achieve its catalytic activity. In the previous section, we outlined the behaviors of various functional groups as the reaction proceeds. Here, we focus on the key unique findings from our simulations and the major differences in comparison to other models:

- two different Mg<sup>2+</sup> ions participate in the reaction, but at different stages, one playing a structural role and another one acting directly as the base catalyst

- the conformation of the G25:U20 pair is flexible and is highly coupled with the reaction coordinate
- the position of C75 and its protonation state change at different stages of the reaction
- the spontaneous rotation of U-1, resulting from the charge environment change after the base catalysis step, leads to an inline conformation poised for the subsequent steps

Note that all of these findings were observed directly from molecular simulations and were not based on any a priori assumptions. As mentioned previously, the purpose of the present work is to identify plausible catalytically active states along the reaction coordinate that can serve as a departure point for further QM/MM investigation to characterize the free energy landscape for the chemical steps of the reaction in order to gain a complete picture of mechanism and estimate of reaction rates. Nonetheless, in order to place the current work into context, it is useful to discuss the general expected time scales involved. The wild-type HDVr has a reaction rate enhancement of nearly 3000-fold, which is in the range of  $\sim 1 \text{ min}^{-1}$ .<sup>96</sup> The breaking of the G25:U20 pair should require free energy similar to base flipping in double-strand RNA, which could be spontaneous in the time scale of nanoseconds<sup>97</sup> for certain sequences. Using high-level quantum calculations, the energy barrier of glycosidic bond rotation DNA bases has been estimated in the range of 3–8 kcal/mol,<sup>98,99</sup> corresponding to half-lives of 20 ps to 80 ns, indicating that the proposed steps of G25 and U-1 rotations should fall into this range of time scale. Together, these proposed mechanistic steps should be feasible in the HDVr chemical reaction.

**Participation of Two Different  $Mg^{2+}$  Ions in the Reaction, One Playing a Structural Role and Another Acting as a Base Catalyst.** Although numerous works have suggested that two distinguishable  $Mg^{2+}$  ions and/or metal binding sites may be involved in the HDVr reaction,<sup>23,32,43,46,65,66,70,73</sup> most recent mechanistic models have focused heavily on “the catalytically critical divalent metal ion”.<sup>36,42,46,65,66,68–72</sup> Less conventionally, our simulation results suggest two  $Mg^{2+}$  ions involved at different binding sites and *at different points along the reaction path*. The *B-site  $Mg^{2+}$ , seen in several crystal structures<sup>58</sup> and inferred from mutational studies,<sup>45,46</sup>* is stable in our simulations when C75 is neutral but is observed to dissociate when C75 is protonated. The most straightforward interpretation of these results is that the B-site  $Mg^{2+}$  likely plays a structural role to promote an active site conformation that facilitates C75 protonation, but this protonation is correlated with release of  $Mg^{2+}$  from this site.

Subsequently, another  $Mg^{2+}$  ion enters the C-site and, *acting as a base catalyst*, facilitates deprotonation of the U-1:O2' nucleophile. This ion also leaves the active site, but only after U-1 rotates to form an inline conformation and the reaction conditions are set. This picture is consistent with the experimental conclusion that there is at least one structural divalent metal ion that does not participate in catalysis and that  $Mg^{2+}$  speeds up the reaction  $\sim 125$ -fold through purely structural roles (the B-site  $Mg^{2+}$ ).<sup>32</sup> Consequently, our model predicts that weakened B-site binding will result in a decrease in enzyme activity but will not alter the reaction pH profile if the conformational change to active state is not rate-limiting. However, absence of the C-site metal ion, the proposed base catalyst, will alter the pH profile and decrease enzyme activity in a high pH range but not at low pH.

**Thio Substitution Studies.** Previous thio substitution studies showed little or no thio effects on the HDVr reaction or a rescue effect from  $Mn^{2+}$ .<sup>75,76</sup> Nevertheless, more recent studies show thio substitution effects in the background of both mono- and divalent ions and an “unconventional” rescue effect by  $Cd^{2+}$  involving suppression of the unmodified ribozyme rate constant.<sup>71,72</sup>

In our model, the *pro-R<sub>p</sub>* oxygen is also important, as it has a water-mediated contact with the B-site  $Mg^{2+}$ , consistent with experimental evidence.<sup>71,72,75,76</sup> In this picture thio substitution of the *pro-R<sub>p</sub>* oxygen will not change the reaction pH profile when  $Mg^{2+}$  is present, consistent with the reported experiments.<sup>71</sup> This thio effect will be largely metal ion independent, since there is no direct binding of the *pro-R<sub>p</sub>* oxygen to the metal ion. Hence whether  $Cd^{2+}$  is present or not, the reaction rate of the *pro-R<sub>p</sub>* thio substitution will be similar. This will lead to the “unconventional rescue effect of  $Cd^{2+}$ ,” mentioned by Thaplyal et al.<sup>71</sup>

**G(1,2)-7DG Mutants.** Chen et al.<sup>65</sup> have studied the effects of 7-deazaguanosine (7DG) substitution at the G1 and G2 positions. This corresponds to the C-site, where our model suggests that a G1-7DG mutation will negatively affect  $Mg^{2+}$  binding. As above, disrupting the C-site binding will alter the pH profile and decrease enzyme activity at the high pH range but not at the low pH range. This was in fact observed at pH 5 and pH 7.<sup>65</sup> The curve shapes of the reaction rate versus  $Mg^{2+}$  concentration are similar for all G(1,2)-7DG mutants at a given pH, indicating that B-site function is not affected. Interestingly, this explanation would require the prediction that contribution of  $Mg^{2+}$  to the chemical step at pH 7 will be around 10-fold, since at pH 7 the rate decrease engendered by G1G2-7DG mutation ( $k_{\max} = 0.62 \text{ min}^{-1}$ ) in comparison to the WT ( $k_{\max} = 7.0 \text{ min}^{-1}$ ) is also 10-fold and will be due to the loss of the C-site  $Mg^{2+}$  (the base catalyst) function. This is qualitatively consistent with the estimates from other studies.<sup>32</sup>

**Rescue of C75Δ Mutants by  $NH_4^+$ .** It has been shown that, in the absence of  $Mg^{2+}$ , only ammonium ions, together with imidazole, rescue inactive C75Δ variants.<sup>72</sup> On the basis of our model, the B-site  $Mg^{2+}$  provides scaffolding to guide C75 toward the active site, where it subsequently becomes protonated. In a purely monovalent ionic environment, such a role will not be filled and the active site of C75Δ may not possess enough space for imidazole to enter. However, the size of  $NH_4^+$  is much larger than other monovalent ions and may be able to function similarly to a hydrated  $Mg^{2+}$  and hence create the necessary space for imidazole to enter. In such a scenario, the pH profile will again be altered as the B-site function is partially restored but not C-site function, consistent with the observed result.<sup>72</sup>

**Anti-Correlation between C75 and  $Mg^{2+}$  Binding.** Several studies have indicated that the shift in the apparent  $pK_a$  of C75 is anti-correlated with increasing  $Mg^{2+}$  concentration.<sup>30,35,36,42,65,70</sup> Here our simulations show that the B-site  $Mg^{2+}$  and protonated C75 repel each other. Increased concentration of  $Mg^{2+}$  will, in principle, increase occupation of all binding sites, but in this scenario anticorrelation with C75 only originates from the B-site. In a study of G1/G2-7DG mutants by Chen et al.,<sup>65</sup> anti-correlation between C75 and  $Mg^{2+}$  was not reported. Nevertheless, on the basis of our model, such anti-correlation should be observed for G1/G2-7DG mutants where only C-site binding will be affected.

**Further Prediction.** According to our model, the phosphoryl transfer and the general acid components of the reaction will be

$Mg^{2+}$ -independent because both B-site and C-site ions are already dissociated from the active site at this point. Recent QM/MM studies,<sup>69</sup> however, have predicted the reaction to be highly  $Mg^{2+}$ -dependent: i.e., the activation free energy differs by more than 8 kcal/mol on comparison of the reaction in the presence and absence of  $Mg^{2+}$ .<sup>69,84</sup> Further experimental evidence may be needed to distinguish them.

**Flexibility of G25:U20 and Coupling of Its Conformation Is Coupled with the Reaction Coordinate. Formation of the G25:U20 Reverse Wobble via B-Site  $Mg^{2+}$  Binding.** The tWW (reverse wobble) form of G25:U20 has been observed in both a precleavage crystal structure<sup>44</sup> and during a recent refitting of precleavage C75U and product structures.<sup>47</sup> Several studies have suggested that G25:N<sub>7</sub> creates a metal binding pocket in the active site;<sup>26,46,58,70,81</sup> however, the tWW form of G25:U20 has never been directly observed along with U-1 in wild-type HDVr, and it is not clear whether G25:U20 is in its tWW form prior to the reaction.

Our simulation results suggest that the tWW form is likely the result of  $Mg^{2+}$  binding at the B-site and that the G25:U20 base pairing could become relatively weak (Table 1 and Figure 3) and therefore unstable in the absence of  $Mg^{2+}$  at the B-site. In fact, simulations without the B-site  $Mg^{2+}$  present show the G25:U20 base pair spontaneously moving toward the tWH conformation (tWW\_RT\_C75° in Table 1). On the other hand, when this ion is present, G25:U20 prefers the tWW conformation due to water-mediated interactions between the ion and G25:N<sub>7</sub>. This observation is consistent with the proposals based on kinetic data of a metal ion dependent conformational change near the cleavage site,<sup>50</sup> the formation of P1.1 assisted by  $Mg^{2+}$ ,<sup>51</sup> as well as the fact that the conformation of G25 was not resolved when metal ions were removed in 1VCS.

**Correlation of the Conformational Switching of the G25:U20 Base Pair with the Protonation of C75 and Recruitment of C-Site  $Mg^{2+}$  Serving as the Base Catalyst.** After C75 is protonated, the B-site  $Mg^{2+}$  will leave and the G25:U20 pair switch to the tWH/cWW conformation. The resulting rearrangement of the active site creates a large space at the C-site and hence recruits another  $Mg^{2+}$  into the C-site. This  $Mg^{2+}$  is at the proper position to serve as a base catalyst. In addition, when the G25:U20 pair is in the tWH/cWW form, it will not provide metal binding ability at the B-site, which allows C75<sup>+</sup> to move toward the active site and act as the general acid. This picture is consistent with the work of Lèvesque et al.,<sup>45</sup> which suggested that the tWH pairing of G25:U20 is formed before the cleavage and this tWH paring is catalytically important. Hence in our model, both the tWW and tWH conformations of G25:U20 are important: the tWW binds to B-site  $Mg^{2+}$  to create the active conformation of the reactant state, while the tWH maintains the active integrity so that the C-site  $Mg^{2+}$  can be recruited after C75 is protonated and B-site  $Mg^{2+}$  leaves.

It has been shown that, although the pair conformation may be flexible, both G25 and U20 are totally conserved across different species, implying that they are critical for the HDVr reaction.<sup>19</sup> Our model demonstrates that both the tWW and tWH conformations are important but for different stages of the reaction, implying that the G25:U20 pair may in fact regulate the progress of the HDVr reaction through its conformation.

**A25:C20 Double Mutant.** From our model, predictions and verification against experimental evidence can be made. In comparison to the wild-type, the A25:C20 double mutant should have almost identical properties and active site microenvironment.

A25:N<sub>7</sub> is capable of providing a similar metal binding pocket for B-site binding. However, A25:C20 has a very different hydrogen bonding pattern in the tWH form and cannot form a stable cWW pairing ("III" in Figure 4) to maintain the active site integrity and to create the C-site to recruit another  $Mg^{2+}$  to serve as a base catalyst. Hence, the A25:C20 double mutant should maintain B-site  $Mg^{2+}$  function but lose C-site function. As a result, the  $Mg^{2+}$  binding affinities for G25:U20 and A25:C20 ought to be similar but the A25:C20 pH profile will be altered. The activity of this mutant at high pH range will be decreased but not at low pH range. In other words, the A25:C20 reaction pH profile will be similar to the reaction without  $Mg^{2+}$ , but only at low pH. A25:C20 should be more active than the reaction without  $Mg^{2+}$ , as the B-site  $Mg^{2+}$  function is still retained. All of these assertions are consistent with known experimental evidence.<sup>45,46</sup>

As for the  $pK_a$  of C75, our model predicts that the anti-correlation relationship between C75 and  $Mg^{2+}$  will still be present in A25:C20 mutants, in contrast to conclusions by Chen et al.<sup>46</sup> and Thaplyal et al.<sup>72</sup> Thaplyal et al.<sup>71</sup> measured the reaction rate of a A25:C20 double mutant. With 10 mM  $Mg^{2+}$  the rate constant is only twice as high at pH 5.6 than at pH 7 ( $k_{obs} = 0.06$  and  $0.03 \text{ min}^{-1}$ , respectively), indicating that the pH profile plateaus in this range in the presence of 10 mM  $Mg^{2+}$ . Hence, the apparent  $pK_a$  of C75 may be near or within this range. On the other hand, with 50 mM  $Mg^{2+}$  the rate constant is ~28 times higher at pH 5.6 than at pH 7 ( $k_{obs} = 0.72$  and  $0.026 \text{ min}^{-1}$ , respectively), indicating that the pH profile has a large negative slope at this range. Hence, the apparent  $pK_a$  of C75 may be shifted significantly downward. We interpret these data as evidence of the anti-correlation between C75 and  $Mg^{2+}$ , implying the B-site binding is retained in the A25:C20 double mutant.

Chemical modifications of G25:U20 which alter B-site binding, formation of the tWW or tWH conformations, or the equilibrium between them (e.g., changing the G25  $\chi$  angle distribution<sup>100</sup>) should also affect the reaction rate and/or the rate pH profile.

**Variance in the Position of C75 and Its Protonation State at Different Stages of the Reaction.** Most simulations of the HDVr reported in the literature begin exclusively with C75 prepositioned in a geometry where it is ready to act as the general acid.<sup>40,66,68,69,71,72,79–83</sup> By contrast, our previous simulations started with the C75 position taken from a C75U mutant trapped in a precleavage state.<sup>42,59</sup> Nonetheless, that work demonstrated the movement of C75 toward the active site when it was protonated and O2' nucleophile O2' was deprotonated. Such trajectories eventually reached a geometry in which C75 was ready to act as the general acid in active sites that mimicked a transition state. The model presented here elaborates this picture further by suggesting that the position of C75 and its protonation state vary at different stages of the reaction. Our model is consistent with the NMR work by Lupták et al.:<sup>37</sup> C75 is presumed to be deprotonated in the reactant and product states and becomes positioned for reaction only along the trajectory from precursor to products.<sup>39</sup>

**Spontaneous Rotation of U-1, Resulting from the Change in Electrostatic Environment after the Base Catalysis Step, Leading to the Inline Conformation.** As mentioned earlier, the inline conformation necessary for the reaction has never been observed in any reported crystal structure to date. Although it has been suggested that there may

**Table 5. Model Summary: Roles of Mg<sup>2+</sup>**

model G <sup>44</sup>	model W <sup>40</sup>	model Y
<u>Initial Structure</u>		
precleavage structure with U-1 modified to deoxynucleotide <sup>44</sup>	precleavage structure with U-1 modified to deoxynucleotide <sup>44</sup>	precleavage C75U structure with C75 mutated back to C
active site conformation modeled on the basis of HHR and in an inline conformation	active site conformation modeled on the basis of HHR and in an inline conformation	active site conformation from the crystal structure and <i>not</i> in an inline conformation
one Mg <sup>2+</sup> ion fixed at B-site	one Mg <sup>2+</sup> ion fixed at B-site	<u>Mg<sup>2+</sup></u>
Mg <sup>2+</sup> ion directly bound to the nonbridging oxygen of the reactive phosphoryl group	Mg <sup>2+</sup> ion indirectly bound (via water) to the nonbridging oxygen of the reactive phosphoryl group	two Mg <sup>2+</sup> ions enter the active sites at different stages: one at B-site, one at C-site
Mg <sup>2+</sup> ion may serve as a Lewis acid or the general base	this Mg <sup>2+</sup> may leave after C75 is protonated (not clear) another Mg <sup>2+</sup> comes in after C75 is protonated (but not at the C-site) and serves as the base catalyst	B-site Mg <sup>2+</sup> is bound (via water) to G25:N7 B-site Mg <sup>2+</sup> stabilizes the reactant state conformation causing shift in C75 pK <sub>a</sub> B-site Mg <sup>2+</sup> binding lost after C75 is protonated
Mg <sup>2+</sup> is near the leaving group all the time and would have a strong effect on the general acid step	not clear	C-site Mg <sup>2+</sup> binds to G1:N7 after B-site Mg <sup>2+</sup> leaves and is positioned to act as the base catalyst and leaves when the reaction reaches the S <sub>N</sub> 2-like transition state
<u>Consequences and Predictions</u>		
Mg <sup>2+</sup> will not have any effect on the general acid step		
<u>Experimental Evidence</u>		
two possible Mg <sup>2+</sup> binding sites <sup>23,43,65,66</sup> and two classes of Mg <sup>2+</sup> ions <sup>25,32,66,67</sup>	Mg <sup>2+</sup> may serve to modulate the pK <sub>a</sub> of C75 <sup>30</sup>	kinetic evidence that at least one structural divalent metal ion that does not participate in catalysis <sup>32</sup>

**Table 6. Model Summary: Thio Effect and C75**

model G <sup>44</sup>	model W <sup>40</sup>	model Y
<u>Thio Effect</u>		
Mg <sup>2+</sup> directly binds to pro-R <sub>p</sub> oxygen all the time	Mg <sup>2+</sup> directly binds to pro-R <sub>p</sub> oxygen before C75 is protonated	B-site Mg <sup>2+</sup> indirectly binds to pro-R <sub>p</sub> oxygen before C75 is protonated, and loses the binding after C75 is protonated
<u>Consequences and Predictions</u>		
thio-R <sub>p</sub> will reduce the probability of forming the active precursor <i>and</i> weaken the general base step	thio-R <sub>p</sub> will reduce the probability of forming the active precursor <i>but</i> not weaken the general base step	thio-R <sub>p</sub> will reduce the probability of forming the active precursor <i>but</i> not weaken the base catalysis step
<u>Experimental Evidence</u>		
pH profile shapes do not change with thio-R <sub>p</sub> substitutions <sup>71</sup>	rescue of thio-R <sub>p</sub> : none with Mn <sup>2+</sup> , <sup>75,76</sup> unconventional with Cd <sup>2+</sup> <sup>71</sup>	
<u>C75</u>		
C75 position is fixed during the reaction, and remains protonated	B-site Mg <sup>2+</sup> stabilizes the reactant state conformation in which C75 pK <sub>a</sub> is shifted upward after C75 is protonated, it recruits another Mg <sup>2+</sup> . (not clear about B-site Mg <sup>2+</sup> ) C75 keeps near the leaving group	B-site Mg <sup>2+</sup> stabilizes the reactant state conformation in which C75 pK <sub>a</sub> is shifted upward when C75 is protonated, it knocks out the B-site Mg <sup>2+</sup> , and recruits the C-site Mg <sup>2+</sup> C75 moves toward the leaving group at the transition state
<u>Consequences and Predictions</u>		
C75 pK <sub>a</sub> will be shifted all the time	C75 pK <sub>a</sub> will only be shifted during the reaction	C75 pK <sub>a</sub> will only be shifted during the reaction
<u>Experimental Evidence</u>		
C75 pK <sub>a</sub> of the reactant and the product shown to be not shifted, observed by NMR <sup>37</sup>	C75 at the reactive position <i>only</i> during the reaction <sup>39</sup>	

be space for U-1 to undergo a rigid rotation to reach the necessary orientation,<sup>42</sup> such a rotation has not yet been directly observed experimentally.<sup>24</sup> Simulations to date have also not reported such a rotation, as they have instead assumed an inline U-1 starting conformation modeled from the HHR.<sup>44</sup> We reported the U-1 rotation toward its inline conformation after the nucleophile O2' is deprotonated.<sup>59</sup>

**Predictions.** If the rotation of U-1 is a necessary step prior to the base catalysis, factors affecting the rotation will also affect the reaction rate. Such alterations might include changing the identity of the -1 position or the sequence and/or length of the upstream strand. Indeed, these have been already observed

experimentally but with different interpretations.<sup>101,102</sup> Chemical modifications altering the rotation energy barrier may be used to test our proposed U-1 motion here. Furthermore, the proposed model predicts that the base catalysis step and the phosphoryl transfer step will be well separated; hence, the base catalysis step should be specific base catalysis. An <sup>18</sup>k<sub>nuc</sub> experiment on the nucleophile O2' would characterize its protonation state along the reaction pathway and verify this prediction.<sup>103,104</sup>

**Comparison to Other Proposed Models.** This proposed dynamic model differs significantly from other proposals. A recent model, here denoted as model G (based on studies by the original proponent, Golden<sup>58</sup>) (Tables 5–7), has been

**Table 7. Model Summary: Inline Conformation**

model G <sup>44</sup>	model W <sup>40</sup>	model Y
U-1 modeled in an inline conformation initially and not changed	U-1 modeled in an inline conformation initially inline conformation destabilized when C75 is deprotonated	U-1 conformation taken from precleavage crystal structure (PDBID: 1VC7) and is not in any inline conformation initially U-1 rotates to form an inline conformation after the nucleophile is deprotonated (observed from simulation)
U-1 should be in the inline conformation in the reactant state	U-1 does not need to rotate to form the inline conformation	U-1 should <i>not</i> be in the inline conformation in the reactant state
upstream nucleotides will not affect the inline conformation	C75 protonation will stabilize the inline conformation upstream nucleotides will not affect the inline conformation	C75 protonation will cause the rotation of U-1 and thus the formation of the inline conformation upstream nucleotides will affect the inline conformation
<b>Consequences and Predictions</b>		
no experimental structure shows U-1 in an inline conformation so far <sup>24</sup>		
U-1 conformation not resolved in a precleavage structure <sup>44</sup>		
<b>Experimental Evidence</b>		

established on the basis of a precleavage structure with a modeled active site and molecular simulations.<sup>44,71,81</sup> It provides very different views of the roles of both the B-site and the C-site Mg<sup>2+</sup> ions,<sup>26,46,66</sup> as well as the effect of an A25:C20 double mutant.<sup>46</sup> Model G is more or less static as the active site arrangement is maintained from beginning of the base catalysis step to the very end of the general acid step. The inline conformation is enforced from the homologous model and U-1 conformation change (rotation) is not needed in this model.

Model G has been further extended with molecular dynamics simulations, and it has been suggested that it may be a reactive intermediate trapped by low pH. A set of scenarios of possible dynamic paths describing the interplay between the U-1 and G1 conformations, and the protonated C75, as well as the possible participation of the second Mg<sup>2+</sup>, were proposed and here are denoted as model W (Tables 5–7).<sup>40</sup>

Conversely, our model, denoted as model Y (Tables 5 to 7), begins with a crystal structure of the HDVr with U-1 completely resolved. Simulations were performed with different conditions (various G25:U20 conformations, Mg<sup>2+</sup> binding modes, C75 protonation states, and different reaction stages), and a dynamic picture of the HDVr reaction is deduced from these simulations and experimental evidence. Hence, this proposed dynamic model significantly differs from other proposed models and further experimental efforts will be needed to distinguish all possible hypothesized models.

## CONCLUSION

A set of long-term molecular dynamics simulations of the HDVr at different stages along the reaction path have been performed. Simulation results predict a dynamic picture of the HDVr reaction mechanism whereby switches among three possible G25:U20 base pairing conformations play important roles in forming proper metal binding environments at different stages of the reaction. Two Mg<sup>2+</sup> ions are coupled with this switching in a sequential fashion. The first ion plays a structural role by inducing a base pair flip necessary to obtain the catalytic fold in which C75 moves toward the scissile phosphate in the active site. The second ion is poised to possibly play a chemical role by acting as a base catalyst through a bound water. Both ions are directly involved in the reaction but at different stages. This model offers an alternate mechanistic interpretation of a broad range of currently available experimental data. Several experimentally testable predictions are made that can be used

to further lend credence to, or else refute, the model. Spectroscopic signals such as NMR chemical shifts<sup>105</sup> to monitor the syn/anti conformation of G25 or chemical modification of G25 that leads to altered syn/anti equilibria will be particularly valuable.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acscatal.5b02158.

Additional computational details ([PDF](#))

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### Notes

The authors declare no competing financial interest.

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## REFERENCES

- (1) Quin, M. B.; Schmidt-Dannert, C. *ACS Catal.* **2011**, *1*, 1017–1021.
- (2) Choi, J.-M.; Han, S.-S.; Kim, H.-S. *Biotechnol. Adv.* **2015**, *33*, 1443.
- (3) Garcia-Viloca, M.; Gao, J.; Karplus, M.; Truhlar, D. G. *Science* **2004**, *303*, 186–195.
- (4) Kruger, K.; Grabowski, P.; Zaug, A.; Sands, J.; Gottschling, D.; Cech, T. *Cell* **1982**, *31*, 147–157.
- (5) Latham, J. A.; Cech, T. R. *Science* **1989**, *245*, 276–282.
- (6) Zaug, A. J.; Cech, T. R. *Science* **1986**, *231*, 470–475.
- (7) Guerrier-Takada, C.; Haydock, K.; Allen, L.; Altman, S. *Biochemistry* **1986**, *25*, 1509–1515.
- (8) Haseloff, J.; Gerlach, W. L. *Nature* **1988**, *334*, 585–591.
- (9) Penchovsky, R. *Biotechnol. Adv.* **2014**, *32*, 1015–1027.
- (10) Narlikar, G. J.; Herschlag, D. *Annu. Rev. Biochem.* **1997**, *66*, 19–59.
- (11) Doudna, J. A.; Lorsch, J. R. *Nat. Struct. Mol. Biol.* **2005**, *12*, 395–402.
- (12) Lilley, D. M. J. *Philos. Trans. R. Soc., B* **2011**, *366*, 2910–2917.

- (13) Ward, W. L.; Plakos, K.; DeRose, V. J. *Chem. Rev.* **2014**, *114*, 4318–4342.
- (14) Kuo, M. Y.; Sharmin, L.; Dinter-Gottlieb, G.; Taylor, J. J. *Virology* **1988**, *62*, 4439–4444.
- (15) Sharmin, L.; Kuo, M. Y.; Dinter-Gottlieb, G.; Taylor, J. J. *Virology* **1988**, *62*, 2674–2679.
- (16) Wu, H.-N.; Lin, Y.-J.; Lin, F.-P.; Makino, S.; Chang, M.-F. *Proc. Natl. Acad. Sci. U. S. A.* **1989**, *86*, 1831–1835.
- (17) Lai, M. M. C. *Annu. Rev. Biochem.* **1995**, *64*, 259–286.
- (18) Salehi-Ashtiani, K.; Lupták, A.; Litovchick, A.; Szostak, J. W. *Science* **2006**, *313*, 1788–1792.
- (19) Webb, C.-H. T.; Riccitelli, N. J.; Ruminski, D. J.; Lupták, A. *Science* **2009**, *326*, 953.
- (20) Webb, C.-H. T.; Lupták, A. *RNA Biol.* **2011**, *8*, 719–727.
- (21) Sánchez-Luque, F. J.; López, M. C.; Carreira, P. E.; Alonso, C.; Thomas, M. C. *BMC Genomics* **2014**, *15*, 340.
- (22) Zhu, J. Y. A.; Meyer, I. M. *RNA Biol.* **2015**, *12*, 5–20.
- (23) Riccitelli, N. J.; Delwart, E.; Lupták, A. *Biochemistry* **2014**, *53*, 1616–1626.
- (24) Been, M. D. *Curr. Top. Microbiol. Immunol.* **2006**, *307*, 47–65.
- (25) Fedor, M. J. *Annu. Rev. Biophys.* **2009**, *38*, 271–299.
- (26) Golden, B. L.; Hammes-Schiffer, S.; Carey, P. R.; Bevilacqua, P. C. In *Biophysics of RNA Folding*; Russell, R., Ed.; Springer: New York, 2013; Biophysics for the Life Sciences, Vol. 3, Chapter 8, pp 135–167.
- (27) Doudna, J. A.; Ferré-D'Amare, A. R.; Zhou, K. *Nature* **1998**, *395*, 567–574.
- (28) Perrotta, A. T.; Been, M. D. *Nucleic Acids Res.* **1990**, *18*, 6821–6827.
- (29) Wadkins, T. S.; Perrotta, A. T.; Ferré-D'Amare, A. R.; Doudna, J. A.; Been, M. D. *RNA* **1999**, *5*, 720–727.
- (30) Nakano, S.; Chadalavada, D. M.; Bevilacqua, P. C. *Science* **2000**, *287*, 1493–1497.
- (31) Das, S.; Piccirilli, J. *Nat. Chem. Biol.* **2005**, *1*, 45–52.
- (32) Nakano, S.; Proctor, D. J.; Bevilacqua, P. C. *Biochemistry* **2001**, *40*, 12022–12038.
- (33) Oyelere, A. K.; Kardon, J. R.; Strobel, S. A. *Biochemistry* **2002**, *41*, 3667–3675.
- (34) Das, S.; Piccirilli, J. *Nat. Chem. Biol.* **2005**, *1*, 45–52.
- (35) Gong, B.; Chen, J.-H.; Chase, E.; Chadalavada, D. M.; Yajima, R.; Golden, B. L.; Bevilacqua, P. C.; Carey, P. R. *J. Am. Chem. Soc.* **2007**, *129*, 13335–13342.
- (36) Cerrone-Szakal, A. L.; Siegfried, N. A.; Bevilacqua, P. C. *J. Am. Chem. Soc.* **2008**, *130*, 14504–14520.
- (37) Lupták, A.; Ferré-D'Amare, A. R.; Zhou, K.; Zilm, K. W.; Doudna, J. A. *J. Am. Chem. Soc.* **2001**, *123*, 8447–8452.
- (38) Shih, I.; Been, M. D. *Proc. Natl. Acad. Sci. U. S. A.* **2001**, *98*, 1489–1494.
- (39) Harris, D. A.; Rueda, D.; Walter, N. G. *Biochemistry* **2002**, *41*, 12051–12061.
- (40) Sripathi, K. N.; Tay, W. W.; Banáš, P.; Otyepka, M.; Šponer, J.; Walter, N. G. *RNA* **2014**, *20*, 1112–1128.
- (41) Leontis, N. B.; Westhof, E. *RNA* **2001**, *7*, 499–512.
- (42) Ke, A.; Zhou, K.; Ding, F.; Cate, J. H. D.; Doudna, J. A. *Nature* **2004**, *429*, 201–205.
- (43) Ke, A.; Ding, F.; Batchelor, J. D.; Doudna, J. A. *Structure* **2007**, *15*, 281–287.
- (44) Chen, J.-H.; Yajima, R.; Chadalavada, D. M.; Chase, E.; Bevilacqua, P. C.; Golden, B. L. *Biochemistry* **2010**, *49*, 6508–6518.
- (45) Lévesque, D.; Reymond, C.; Perreault, J.-P. *PLoS One* **2012**, *7*, e40309.
- (46) Chen, J.; Ganguly, A.; Miswan, Z.; Hammes-Schiffer, S.; Bevilacqua, P. C.; Golden, B. L. *Biochemistry* **2013**, *52*, 557–567.
- (47) Kapral, G. J.; Jain, S.; Noeske, J.; Doudna, J. A.; Richardson, D. C.; Richardson, J. S. *Nucleic Acids Res.* **2014**, *42*, 12833–12846.
- (48) Lynch, S. R.; Tinoco, I., Jr *Nucleic Acids Res.* **1998**, *26*, 980–987.
- (49) Brown, T. S.; Chadalavada, D. M.; Bevilacqua, P. C. *J. Mol. Biol.* **2004**, *341*, 695–712.
- (50) Rosenstein, S. P.; Been, M. D. *Biochemistry* **1990**, *29*, 8011–8016.
- (51) Lévesque, D.; Choufani, S.; Perreault, J.-P. *RNA* **2002**, *8*, 464–477.
- (52) Kumar, P. K.; Taira, K.; Nishikawa, S. *Biochemistry* **1994**, *33*, 583–592.
- (53) van Tol, H.; Buzatan, J. M.; Feldstein, P. A.; Eckstein, F.; Bruening, G. *Nucleic Acids Res.* **1990**, *18*, 1971–1975.
- (54) Slim, G.; Gait, M. J. *Nucleic Acids Res.* **1991**, *19*, 1183–1188.
- (55) Takagi, Y.; Warashina, M.; Stec, W. J.; Yoshinari, K.; Taira, K. *Nucleic Acids Res.* **2001**, *29*, 1815–1834.
- (56) Yoshinari, K.; Taira, K. *Nucleic Acids Res.* **2000**, *28*, 1730–1742.
- (57) Stowasser, R.; Usher, D. A. *Bioorg. Chem.* **2002**, *30*, 420–430.
- (58) Golden, B. L. *Biochemistry* **2011**, *50*, 9424–9433.
- (59) Lee, T.-S.; Giambaşu, G. M.; Harris, M. E.; York, D. M. *J. Phys. Chem. Lett.* **2011**, *2*, 2538–2543.
- (60) Pereira, M. J. B.; Harris, D. A.; Rueda, D.; Walter, N. G. *Biochemistry* **2002**, *41*, 730–740.
- (61) Suh, Y. A.; Kumar, P. K.; Taira, K.; Nishikawa, S. *Nucleic Acids Res.* **1993**, *21*, 3277–3280.
- (62) Shih, I.; Been, M. D. *RNA* **1999**, *5*, 1140–1148.
- (63) Shih, I.; Been, M. D. *Annu. Rev. Biochem.* **2002**, *71*, 887–917.
- (64) Nakano, S.; Bevilacqua, P. C. *Biochemistry* **2007**, *46*, 3001–3012.
- (65) Chen, J.-H.; Gong, B.; Bevilacqua, P. C.; Carey, P. R.; Golden, B. L. *Biochemistry* **2009**, *48*, 1498–1507.
- (66) Veeraraghavan, N.; Ganguly, A.; Golden, B. L.; Bevilacqua, P. C.; Hammes-Schiffer, S. *J. Phys. Chem. B* **2011**, *115*, 8346–8357.
- (67) Sigel, R. K. O.; Pyle, A. M. *Chem. Rev.* **2007**, *107*, 97–113.
- (68) Mlynský, V.; Walter, N. G.; Šponer, J.; Otyepka, M.; Banáš, P. *Phys. Chem. Chem. Phys.* **2015**, *17*, 670–679.
- (69) Ganguly, A.; Thaplyal, P.; Rosta, E.; Bevilacqua, P. C.; Hammes-Schiffer, S. *J. Am. Chem. Soc.* **2014**, *136*, 1483–1496.
- (70) Gong, B.; Chen, J.-H.; Bevilacqua, P. C.; Golden, B. L.; Carey, P. R. *Biochemistry* **2009**, *48*, 11961–11970.
- (71) Thaplyal, P.; Ganguly, A.; Golden, B. L.; Hammes-Schiffer, S.; Bevilacqua, P. C. *Biochemistry* **2013**, *52*, 6499–6514.
- (72) Thaplyal, P.; Ganguly, A.; Hammes-Schiffer, S.; Bevilacqua, P. C. *Biochemistry* **2015**, *54*, 2160–2175.
- (73) Gong, B.; Chen, Y.; Christian, E. L.; Chen, J.-H.; Chase, E.; Chadalavada, D. M.; Yajima, R.; Golden, B. L.; Bevilacqua, P. C.; Carey, P. R. *J. Am. Chem. Soc.* **2008**, *130*, 9670–9672.
- (74) Shih, I.; Been, M. *Biochemistry* **2000**, *39*, 9055–9066.
- (75) Fauzi, H.; Kawakami, J.; Nishikawa, F.; Nishikawa, S. *Nucleic Acids Res.* **1997**, *25*, 3124–3130.
- (76) Jeoung, Y. H.; Kumar, P. K.; Suh, Y. A.; Taira, K.; Nishikawa, S. *Nucleic Acids Res.* **1994**, *22*, 3722–3727.
- (77) Perrotta, A. T.; Been, M. D. *Biochemistry* **2006**, *45*, 11357–11365.
- (78) Nakano, S.; Bevilacqua, P. C. *J. Am. Chem. Soc.* **2001**, *123*, 11333–11334.
- (79) Sripathi, K. N.; Banáš, P.; Rélová, K.; Šponer, J.; Otyepka, M.; Walter, N. G. *Phys. Chem. Chem. Phys.* **2015**, *17*, 5887–5900.
- (80) Veeraraghavan, N.; Bevilacqua, P. C.; Hammes-Schiffer, S. *J. Mol. Biol.* **2010**, *402*, 278–291.
- (81) Veeraraghavan, N.; Ganguly, A.; Chen, J.-H.; Bevilacqua, P. C.; Hammes-Schiffer, S.; Golden, B. L. *Biochemistry* **2011**, *50*, 2672–2682.
- (82) Banáš, P.; Jurečka, P.; Walter, N. G.; Šponer, J.; Otyepka, M. *Methods* **2009**, *49*, 202–216.
- (83) Banáš, P.; Rulíšek, L.; Hánová, V.; Svozil, D.; Walter, N. G.; Šponer, J.; Otyepka, M. *J. Phys. Chem. B* **2008**, *112*, 11177–11187.
- (84) Radak, B. K.; Lee, T.-S.; Harris, M. E.; York, D. M. *RNA* **2015**, *21*, 1566–1577.
- (85) Case, D. A.; Pearlman, D. A.; Caldwell, J. W.; Cheatham, T. E., III; Wang, J.; Ross, W. S.; Simmerling, C. L.; Darden, T. A.; Merz, K. M.; Stanton, R. V.; Cheng, A. L.; Vincent, J. J.; Crowley, M.; Tsui, V.; Gohlke, H.; Radmer, R. J.; Duan, Y.; Pitera, J.; Massova, I.; Seibel, G. L.; Singh, U. C.; Weiner, P. K.; Kollman, P. A. AMBER 7; University of California San Francisco: San Francisco, CA, 2002.
- (86) Pearlman, D. A.; Case, D. A.; Caldwell, J. W.; Ross, W. R.; Cheatham, T., III; DeBolt, S.; Ferguson, D.; Seibel, G.; Kollman, P. *Comput. Phys. Commun.* **1995**, *91*, 1–41.

- (87) Pérez, A.; Marchán, I.; Svozil, D.; Sponer, J.; Cheatham, T. E., III; Laughton, C. A.; Orozco, M. *Biophys. J.* **2007**, *92*, 3817–3829.
- (88) Mayaan, E.; Moser, A.; MacKerell, A. D., Jr.; York, D. M. *J. Comput. Chem.* **2007**, *28*, 495–507.
- (89) Cerrone-Szakal, A. L.; Chadalavada, D. M.; Golden, B. L.; Bevilacqua, P. C. *RNA* **2008**, *14*, 1746–1760.
- (90) Nakano, S.; Cerrone, A. L.; Bevilacqua, P. C. *Biochemistry* **2003**, *42*, 2982–2994.
- (91) Nishikawa, F.; Shirai, M.; Nishikawa, S. *Eur. J. Biochem.* **2002**, *269*, 5792–5803.
- (92) Lee, T.-S.; Silva-Lopez, C.; Martick, M.; Scott, W. G.; York, D. M. *J. Chem. Theory Comput.* **2007**, *3*, 325–327.
- (93) Lee, T.-S.; Silva Lopez, C.; Giambasu, G. M.; Martick, M.; Scott, W. G.; York, D. M. *J. Am. Chem. Soc.* **2008**, *130*, 3053–3064.
- (94) Lee, T.-S.; Giambacu, G. M.; Sosa, C. P.; Martick, M.; Scott, W. G.; York, D. M. *J. Mol. Biol.* **2009**, *388*, 195–206.
- (95) Wong, K.-Y.; Lee, T.-S.; York, D. M. *J. Chem. Theory Comput.* **2011**, *7*, 1–3.
- (96) Chadalavada, D. M.; Cerrone-Szakal, A. L.; Bevilacqua, P. C. *RNA* **2007**, *13*, 2189–2201.
- (97) Pan, Y.; MacKerell, A. D., Jr. *Nucleic Acids Res.* **2003**, *31*, 7131–7140.
- (98) Foloppe, N.; Hartmann, B.; Nilsson, L.; MacKerell, A. D., Jr. *Biophys. J.* **2002**, *82*, 1554–1569.
- (99) Foloppe, N.; Nilsson, L. *J. Phys. Chem. B* **2005**, *109*, 9119–9131.
- (100) Sokoloski, J. E.; Godfrey, S. A.; Dombrowski, S. E.; Bevilacqua, P. C. *RNA* **2011**, *17*, 1775–1787.
- (101) Chadalavada, D. M.; Knudsen, S. M.; Nakano, S.-i.; Bevilacqua, P. C. *J. Mol. Biol.* **2000**, *301*, 349–367.
- (102) Perrotta, A. T.; Been, M. D. *Biochemistry* **2007**, *46*, 5124–5130.
- (103) Humphry, T.; Iyer, S.; Iranzo, O.; Morrow, J. R.; Richard, J. P.; Paneth, P.; Hengge, A. C. *J. Am. Chem. Soc.* **2008**, *130*, 17858–17866.
- (104) Harris, M. E.; Dai, Q.; Gu, H.; Kellerman, D. L.; Piccirilli, J. A.; Anderson, V. E. *J. Am. Chem. Soc.* **2010**, *132*, 11613–11621.
- (105) Wijmenga, S. S.; van Buuren, B. N. *Prog. Nucl. Magn. Reson. Spectrosc.* **1998**, *32*, 287–387.